A contribution to the knowledge of family Chalcididae (Hymenoptera: Chalcidoidea) in Iran

H. Lotfalizadeh^{1&*}, E. Ebrahimi² and G. Delvare³

1. Department of Plant Protection, Agricultural and Natural Resources Research of East-Azerbaijan, Tabriz, Iran, 2. Insect Taxonomy Research Department, Iranian Research Institute of Plant Protection, Tehran, P.O. B. 19395-1454, Iran, 3. UMR INRA CIRAD IRD AGROM - Centre de Biologie et de Gestion des Populations (CBGP), Campus International de Baillarguet, 34398 Montpellier Cedex 5, France.

*Corresponding author, E-mail: lotfalizadeh@gmail.com

Abstract

In the course of this study, the number of recorded chalcidids (Hym.: Chalcididae) from Iran increased to 42 species (including one unidentified species of the genus Lasiochalcidia Masi), of which fifteen species and four genera are new to Iranian chalcidid fauna. The family Chalcididae is repredented by three subfamilies in Iran as follows: Chalcidinae (16 species), Dirhininae (two species), and Haltichellinae (24 species). The genus Varzobia Nikol'skaya is here synonymized with Tanycoryphus Cameron and the type species transferred to that genus as Tanycoryphus tibialis (Nikol'skaya, 1960) comb. n. The species Lasiochalcidia indescripta Bouček, 1952 stat. rev. is removed from synonymy with L. guineensis (Steffan, 1951). The validity of the species Peltochalcidia ferrierei Steffan is restored, being removed from junior synonymy with P. benoisti Steffan, and transferred to the genus Psilochalcis Kieffer. The species Peltochalcidia oranensis Bouček, 1952 is here synonymized with P. benoisti. Complementary information on the biology, morphology and distribution of the species as well as a preliminary list of the Iranian chalcidid species are presented.

Key Words: Chalcididae, new combination, new records, fauna, Iran

چکیده

در طی این تحقیق، تعداد گونههای گزارش شده از زنبورهای خانواده ی Chalcididae از ایران به ٤٢ گونه (شامل یک گونه ی شناسایی نشده از جنس Lasiochalcidia Masi) افزایش پیدا کرد که از این تعداد، ۱۵ گونه و چهار جنس برای اولین کونهی شناسایی نشده از جنس Chalcididae در ایران با سه زیرخانواده به شرح زیر شناخته می شود: Paticididae بار از ایران گزارش می شوند. خانوادهی Chalcididae (با ۲۶ گونه). در این بررسی، جنس Dirhininae (با دو گونه) و Haltichellinae (با ۲۶ گونه). در این بررسی، جنس Dirhininae ایر Varzobia Nikol'skaya با کونهی در این بررسی، جنس اخیر متقال شد، Tanycoryphus tibialis با تعیونیمی با جنس اخیر متقال شد، (Nikol'skaya, 1960) comb. n. گونهی المعنونیمی با گونهی Lasiochalcidia indescripta Bouček, 1952 stat. rev. گونهی گونهی Peltochalcidia ferrierei Steffan به عنوان یک گونهی گونهی P. benoisti Steffan بحنوان یک گونهی معتبر به جنس به P. benoisti Steffan با گونهی او از سینونیمی با گونهی گونههای موجود در ایران ارائه شده است. روزگان کلیدی: Peltochalcidia oranensis Bouček, 1952 در ایران ارائه شده است. و پراکنش هرکدام از گونهها و همچنین فهرست اولیهی گونههای موجود در ایران ارائه شده است. و واژگان کلیدی: Chalcididae.

Introduction

The family Chalcididae (Hymenoptera: Chalcidoidea) is a moderate sized family within the superfemily Chalcidoidea, including over 1500 species and 90 genera worldwide (Noyes, 2011). The family is widely distributed but is known to be more diversified in the tropical lowlands (Delvare & Bouček, 1992). Chalcidids are mostly parasitoids, including a minority

of hyperparasitoids on a wide range of holometabolous insect hosts such as Lepidoptera, Diptera, Hymenoptera, Coleoptera, Neuroptera and Strepsiptera as the chalcidid adults always emerge from the pupae, whenever the oviposition occurs (Hanna, 1934; Silvestri, 1943; Bouček, 1952; Steffan, 1959b; Habu, 1960; Narendran, 1986; Delvare & Bouček, 1992). Some species such as *Brachymeria tibialis* (Walker) are economically important for their role as parasitoids of destructive lepidopterous pests (Adeli, 1975).

The following five subfamilies are now recognized within the family: Chalcidinae, Dirhininae, Epitraninae, Haltichellinae and Smicromorphinae (Bouček, 1988; Delvare, 2004; Noyes, 2011). Recent phylogenetic studies, using molecular data (Munro *et al.*, 2011) or combined morphological and molecular data, have been conducted in the context of the Hymenoptera Tree of Life project (Heraty, 2003).

The family Chalcididae is easily recognizable by the combination of following characters: body robust mostly black and/or red with yellow markings or heavily sclerotized with coarsely punctured head and mesosoma (figs 1 & 4). Hind femur enlarged, ventrally toothed or serrulate (fig. 5); hind tibia arched to accommodate femur. Prepectus smaller than tegula, gena carinate posteriorly, propodeum mostly areolate (fig. 4), mesepisternum with a ventral shelf delimited anteriorly by epicnemial carina. Some of these characters are separately found in other families, such as Eurytomidae, Leucospidae and Torymidae but never in combination.

The phylogeny of the family was revised by Wijesekara (1997), using 34 morphological characters of 29 taxa. Bouček (1988) proposed a new classification for the family and provided a key to the subfamilies and genera of Australasian region. Bouček (1952, 1956, 1965, 1972, 1977) and Steffan (1950, 1951a, 1951b, 1951d, 1951e, 1953, 1957, 1959a, 1959b, 1976) extensively revised the West-Palaearctic species of Chalcididae. Nikol'skaya (1952, 1960) and Habu (1960) studied the species fauna of the former USSR and Japan respectively. Monographs on chalcidid species were presented by Narendran (1989) and Delvare & Bouček (1992) for the Oriental region and the New World respectively.

The studies on Iranian chalcidid fauna dates back to 1924, when Masi (1924) described two new species from Iran. Later, Bouček (1952, 1956) and Nikol'skaya (1952, 1960) added six new species and Haeselbarth (1983) listed the Iranian chalcidid species. The family is still poorly known in Iran, being represented only with 16 species in the Universal Chalcidoidea Database (Noyes, 2011), although three new species of economic importance have been recently described (Delvare *et al.*, 2011). The biological and ecological notes on several

Iranian species have been published by Askari & Copple (1977), Sepehr & Abai (1989), Maadani & Ladonni (1993), Dabiri *et al.* (1998) and Sabahi (2000).

Materials and methods

The Hayk Mirzayans Insect Museum (HMIM), Tehran, and additional samples used as the basis of this work. Different collecting techniques such as sweeping, Malaise trap and rearing of hosts were applied during this study. The identifications were made possible using Bouček (1952, 1956, 1988), Delvare *et al.* (1999), Habu (1960), Steffan (1950, 1951a, 1951b, 1951c, 1951d, 1951e, 1953, 1957, 1959a, 1959b, 1976). The third author (GD) examined the types of the species described by Förster (1859), Walker (1871), Masi (1931), Bouček (1948, 1952, 1956, 1972), Steffan (1948, 1951b, 1951c, 1951e), Nikols'kaya (1952, 1960) and Nikol'skaya & Kyao (1954), housed in NHMW, BMNH, MSNG, NHMP, MNHN and ZISP, respectively (see below).

An Iranian list of chalcidid species and their hosts is given (table 1). In diagnosis sections, "mandibular formula 2.3" means the left mandible has 2 teeth, while the right one bears 3 teeth.

Acronyms and institutions

BMNH	Natural History Museum, London, UK
HMIM	Hayk Mirzayans Insect Museum, Tehran, Iran
MSNG	Museo Civica di Storia naturale di Genova, Italy
MNHN	Museum national d'Histoire naturelle de Paris, France
NHMP	Natural History Museum, Praha, Czech Republic
NHMW	Naturhistorisches Museum Wien, Austria
ZISP	Zoological Institute St-Petersburg Russia

Abbreviations

F Funicular segments M Marginal vein PM Postmarginal vein

OOL Distance between inner margin of compound eyes and lateral ocelli

ST Stigmal vein T Gastral tergites

I- Subfamily Chalcidinae

Iranian chalcidines are represented by following species in four genera: *Brachymeria* Westwood, *Chalcis* Fabricius, *Cratocentrus* Cameron and *Trigonura* Sichel.

Brachymeria ceratoniae Delvare, 2011

Material examined – Holotype ♀: Iran: Isfahan, Kashan, viii.2009, ex: *Apomyelois ceratoniae* (Zeller) (Lep.: Pyralidae) on *Ceratonia siliqua* (L. Talaee).

Diagnosis – *Brachymeria ceratoniae* is very close to *B. confalonierii* Masi and its diagnostic characters are quoted in detail by Delvare *et al.* (2011).

Distribution – The species has been recently described by Delvare *et al.* (2011) from Iran as a parasitoid of the carob moth.

Brachymeria excarinata Gahan, 1925

Material examined – Iran: Azarbaijan-e Sharghi, Khosroshahr, 1346 m, N 37° 58′ 28″ & E 46° 02′ 55″, 10.x-6.xi.2007, Malaise trap, (H. Lotfalizadeh), 1 ♀. Tehran, Karaj, 2 Km to Mohammadshahr, 25.viii.2006, ex: *Plutella xylostella* on *Brassica oleraceae*, (A. Golizadeh), 1♀.

Diagnosis – Body and tegula dark. Head with preorbital but without postorbital carina. Mandibles 2.3 with sharp teeth of equal length. F2-F5 slightly transverse. Mesonotum densely punctured. Frenal lobes not expanded, scutellum rounded at apex. Hind femur dull, with engraved network between the dense punctulation. Hind tibia dark with subbasal and dorsoapical pale spots, often reduced. Apical truncature of tibia distinctly emarginate. Gaster slightly shorter than mesosoma. T1 smooth dorsally, T2 punctulate.

Distribution – The species is known from Papua New Guinea and Japan (Joseph *et al.*, 1973), Egypt and Cameroon (Risbec, 1956; Descamps, 1956).

Remarks – *Brachymeria excarinata* is one of the smallest species of the genus and a natural enemy of an important pest of cruciferous crops, the diamondback moth *Plutella xylostella* (L.) (Lep.: Plutellidae). It is otherwise quoted from several families of moths such as Arctiidae, Crambidae, Gelechiidae, Hesperiidae, Noctuidae, Oecophoridae, Pyralidae, Tortricidae (Habu, 1960; Joy *et al.*, 1972; Joseph *et al.*, 1973; Gubbaiah, 1984; Narendran, 1986) and the beetle family of Chrysomelidae (Narendran, 1989). In some cases it was reported as a hyperparasitoid (Descamps, 1956; Risbec, 1956; Joseph *et al.*, 1973). This small form was described as the subspecies *plutellae* Joseph *et al.*, 1972, which was subsequently synonymized with the nominal species by Narendran (1989). The same author also synonymized *B. apantelesi* Risbec with *B. excarinata* without having access to its type series. This synonymy remains doubtful and needs further studies.

Brachymeria femorata (Panzer, 1801)

Material examined – Iran: Ardebil, Moghan, ix.1985, (Damanabi), $1 \circlearrowleft$. Azarbaijan-e Sharghi, Tabriz, iii.1987, (Argand), $1 \circlearrowleft$. Fars, 34 Km north of Nourabad, 6.viii.1975, (M. Abai), $1 \circlearrowleft$. Fars, Kuhmareh, Nodan, 1.xi.1989, (M. Abai), $1 \circlearrowleft$. Kerman, Jiroft - Bam Agriculture Research Centre, summer 1998, ex: pupa of *Heliothis* sp., v.1998, (Mohajeri), $3 \circlearrowleft$. Khuzestan, Andimeshk, Bidrubeh, Galerizeh, 500 m, 30.iv.2001, (E. Gilasian, A. Hajesmailian and M. Mofidi-Neyestanak), $1 \circlearrowleft$. Tehran, Evin, 17.vii.1978, (M. Abai), $1 \circlearrowleft$.

Diagnosis – This species belongs to the same nominal species group in which the body is partly red, mandibular formula 2.2, teeth being acute and of same length, postorbital carinae absent or obliterated, scutellum quite convex, strongly bilobed at apex and densely setose, epicnemial carina with two elevated submedian lobes in front of ventral shelf on mesepisternum, hind tibia slightly emarginate on apical truncation, first gastral tergite dorsally smooth, second and following densely setose. In *B. femorata* the body is black – partly red in some southern specimens – the pale spots on tegula and legs are bright yellow, postorbital carinae present, tibiae entirely and hind femur at both ends yellow; femur otherwise densely punctured, gaster shortly acute. The extent of the black part on the disk of the hind femur is variable and led to the description of several forms (Habu, 1960) which were all found in the material from Iran.

Distribution – This species is distributed from northwest to southwest of Iran and first reported from Iran by Davatchi & Chodjai (1968). Bouček (1956) and Steffan (1959b) mentioned this species as a common species in the Mediterranean region as a pupal parasitoid of various butterflies, mostly Pieridae. It is widely distributed in the Old World, from western Europe to Indonesia (Noyes, 2011).

Brachymeria lasus (Walker, 1841)

Material examined – Iran: Fars, Kazerun, Nowdan, Kuhmareh, 26.vi.1975, (M. Abai), $1 \subsetneq$. Fars, Shiraz - Arsanjan, 21.ix.1996, (M. Abai), $1 \subsetneq$. Kerman, Rafsanjan, 18.x.1974, on *Tamarix*, $1 \subsetneq$ and $1 \circlearrowleft$. No label, $1 \subsetneq$.

Diagnosis – This species belongs to the *annulata* species group with mandibular formula 2.3, teeth acute, postorbital carinae generally present, epicnemial carina forms ventrally raised two submedian lobes in front of the ventral shelf on mesepisternum, hind coxa generally bears a tooth or tubercle in female, apical truncation of hind tibia straight to hardly emarginate. In *B. euploae* frons with dense and thick setation masking integument and

frenal carina moderately expanded, scutellum subtruncate at apex, first gastral tergite smooth and second has very small piliferous points and faint punctulation. Hind tibia pale yellow with a short basal spot and dark stripe along the ventral margin.

Distribution – *Brachymeria lasus* is found in the Palaearctic, Oriental and Australasian regions (Habu, 1960; Joseph *et al.*, 1973; Bouček, 1988; Narendran, 1989) as a parasitoid or sometimes hyperparasitoid of numerous Lepidoptera species (Habu, 1960). It is first reported from Iran.

Brachymeria minuta (Linnaeus, 1767)

Material examined – Iran: Kohkiloye-Boyer-Ahmad, Bahram-Beighi, summer 1994, ex: *Lymanteria dispar* (Lep.: Lymanteridae), $3 \subsetneq \varphi$ and $1 \circlearrowleft$. Fars, Kazerun, Nowdan, Kuhmareh, 8.xi.1989, (M. Abai), $1 \subsetneq$. Sistan-Baluchestan, Sarbaz, vii.1970, (Salavatian), $1 \subsetneq$.

Diagnosis – *Brachymeria minuta* gives its name to a special species group whose mandibles are long, both 2-toothed with blunt teeth and lower tooth longer than upper; head quite transverse in frontal view with distance separating the eyes always greater than their own height; frenal lobes generally well expanded on scutellum that usually emarginate posteriorly and bilobed. The epicnemial carina without submedian ventral lobes on mesepisternum; apical truncation of hind tibia distinctly emarginate. Pale parts bright yellow, scutellum moderately convex and emarginate at apex, hind femur broad, finely punctured, without alutaceous network.

Distribution – Bouček (1952) reported this species from Iran (Kurdistan, Badawa Orbil) (possibly Iraq) and Nikol'skaya (1952) mentioned it from northern part of Iran. The species is widely distributed from the Australasian to the western Palaearctic regions (Bouček, 1952, 1988; Narendran, 1989) including north Africa (Noyes, 2011).

Hosts – This species is parasitic on dipteran Muscoidea especially Muscidae developing on feces. The adults can be seen flying around excrements to find their hosts (GD, personal observation). The reports concerning Lepidoptera evidently concerns individuals developing as hyperparasitoids at the expense of parasitic Diptera such as Tachinidae and Sarcophagidae (Steffan, 1959b; Habu, 1960; Bouček, 1977; Andriescu, 1988).

Brachymeria obtusata (Förster, 1859)

Material examined – Iran: Mazandaran, Noshahr, 25.viii.1987, (M. Abai), $4 \subsetneq Q$. M. Abai personal collection, not labelled, $2 \subsetneq Q$.

Diagnosis – This species belongs to the *minuta* species group for which a diagnosis is given above. It can be separated from other Palaearctic species by the following combination of characters: scutellum never or slightly convex, moderately emarginate at apex; hind femur broad, ×1.7-1.75 as long as wide, densely punctured, mostly red on disk between black basal and yellow apical spots in female, teeth on ventral margin of femur closely set; light parts on tegula and legs pale yellow.

Distribution – *Brachymeria obtusata* is widely reported from north Africa, west Europe and Japan (Noyes, 2011).

Hosts – Although this species is common and widely distributed in southern parts of Palaearctic, very little is known about its biology. Györfi (1939) recorded this species from *Eristalis tenax* (Linnaeus) (Syrphidae) in Hungary. Steffan (1959b) examined a female reared from a Diptera, probably *Sarcophaga* sp. (Sarcophagidae), having developed within the shell of a snail (*Helix* sp.).

Remarks – This species was originally described as *Chalcis vicina* (Walker, 1834). Later, the name changed to *Brachymeria walkeri* by Dalla Torre (1898) because of its primary homonymy with *Chalcis vicina* Fonscolombe (1832). *Chalcis obtusata* Förster, 1859 was recognized as the same species as *Chalcis vicina* Walker by Bouček (1952) and Graham (1992) finely fixed the species through the designation of a neotype.

Brachymeria persica (Masi, 1924)

Distribution – This species was collected from central region of Iran (Kerman) by G. Doria (Masi, 1924, 1951).

Remarks – Masi (1924) named this species as *Chalcis persica* from Iran based on a unique female specimen. It was later transferred into the genus, *Brachymeria* (Masi, 1951). From comparison with *B. tibialis* and *B. femorata*, Masi (1924) concluded that this species belongs to another species group.

Brachymeria rufigaster (Masi, 1924) comb. n.

Chalcis rufigaster Masi, 1924

Distribution – This species was originally described based on a unique male specimen from Kerman, Iran collected by G. Doria. Its type is deposited in the Museum of Genoa (Masi, 1924).

Remarks – Masi (1924) included this species in the *minuta* species group.

Brachymeria rugulosa (Förster, 1859)

Material examined – Syntypes: the two remaining syntypes in the Förster's collection (NHMW). Other material: Iran: Yazd, Ardakan, Samad-Abad, summer 1991, ex: *Apomyelois ceratoniae* (Zeller) (Lep.: Pyralidae), (Ahmadian), $1 \$

Diagnosis – Mandibular formula 2.3, teeth acute, postorbital carinae absent, mesonotum fairly densely punctured but interspaces not raised, scutellum broadly rounded on apical margin, epicnemial carina not raised into submedian lobes in front of ventral shelf of mesepisternum, hind femur moderately densely punctulate, shiny between points, hind tibia testaceous at base with apicodorsal spot, gaster oval acute, T1 smooth, T2 finely punctulate dorsally.

Distribution – The species is distributed from Europe to Turkmenistan (Noyes, 2008). **Hosts** – *Brachymeria rugulosa* attacks various small moths (microlepidoptera) and the

specimen examined from Iran was reared from the carob moth on pomegranate.

Brachymeria tibialis (Walker, 1834)

Diagnosis – This species also belongs to the *annulata* species group, diagnosed above. Postorbital carinae often irregular or incomplete, setation on frons is not especially dense and does not mask the integument, scutellum hardly emarginate at apex, hind coxal tooth acute in female, relevant femur with variable broad apical pale spot, hind tibia yellow with a more or less expanded median stripe, gaster moderately long, T1 smooth, T2 with dorsal piliferous punctures at base together with fine punctulation.

Askew & Shaw (2001) showed a few slight morphological differences between the specimens reared from *Zygaena* spp. and those reared from other Lepidoptera. They argued that primary parasitoids of *Zygaenidae* must avoid the cyanogenic glucosides found in the body content of these moths. This contradicts the broad spectrum of hosts of *B. tibialis*. It is therefore possible that another species is involved and a molecular study is presently carried out at Montpellier to solve the problem.

Distribution – This common species was frequently reported from Iran as its junior synonym, *B. intermedia* (Nees) (Adeli, 1975; Tremewan, 1976; Haeselbarth, 1983; Abai & Faseli, 1986; Fry, 1989; Askew & Shaw, 2001; Nikdel *et al.*, 2004). The material examined shows that this species is widely distributed in Iran from north to south. *B. tibialis* is otherwise widely distributed in the Oriental and Palaearctic regions (Noyes, 2011).

Hosts – Noyes (2011) quotes about 70 insect hosts, including Lepidoptera and tachinids. The species is therefore a facultative hyperparasitoid. It was introduced to North America to control the gypsy moth at the end of the nineteen century. In Iran, Abai (1976) reared this species from the lymantriids *Euproctis melania* Staudinger and *Leucoma wiltshirei* Collenette in southwest of Iran. Abai & Faseli (1986) mentioned the fig tree defoliator *Ocnerogyia amanda* Staudinger as another lymantriid host. Tremewan in 1997 reared a female specimen from *Zygaena haberhaueri* Lederer (Lep.: Zygaenidae) in Mazandaran province (Askew & Shaw, 2001). *Euproctis chrysorrhoea* (L.) was also quoted as its host in northwest (Nikdel *et al.*, 2004), *Parocneria terebynthina* (Staudinger) (Lep.: Lymantriidae) in central part (Kerman, Jiroft) and south of Iran (Fars province) (Sepehr & Abai, 1989; Sabahi, 2000). Sabahi (2000) reported 40.2% parasitism of *B. tibialis* on *P. terebynthina* pupae in the Fars province and the mean parasitism was 25.3%.

Brachymeria vitripennis (Förster, 1859)

Material examined – Holotype ♂: from the Förster's collection (NHMW). Other material: Iran: Azarbaijan-e Sharaghi, Basmenj, xii.1995, (Damanabi), 1 ♂.

Diagnosis – *Brachymeria vitripennis* belongs to the *inermis* species group which includes parasitoids of Cassidinae. Mandibular formula 2.2, teeth sharp and subequal, postorbital carinae absent, malar space large, at least ×0.75 eye height, faint preorbital carinae present, scutellum apically rounded, epicnemial carina not raised into submedian lobes in front of ventral shelf, apical truncation of hind tibia distinctly emarginate, gaster short and ovoid, T1 smooth, T2 with faint punctulation. Puncturation of the mesonotum dense, including subpolygonal points, hind femur sparsely and weakly punctulate, smooth between punctures, hind tibia with a basal pale spot.

Distribution – The species is known from southern Europe to Central Asia (Noyes, 2011). It has further eastern distribution than *Brachymeria inermis* (Fonscolombe), another parasitoid of *Cassida* spp. This is the first record of *B. vitripennis* from Iran.

Hosts – *Brachymeria vitripennis* is parasitic on *Cassida* spp. (Col.: Chrysomelidae: Cassidinae) (Györfi, 1942; Menozzi, 1947; Györfi, 1962).

Chalcis biguttata Spinola, 1808

Material examined – Iran: Mazandaran, Behshahr - Ghalughah, Lemrask, 3.vii.2004, (Zarghami), 1 ♂.

Distribution – The species has been recorded from Europe to Caucasus and Iran (Bouček, 1952, 1966; Nikol'skaya, 1952, 1960).

Remarks – A male specimen of *C. biguttata* without hind legs is placed in the HMIM collection was identified according to its finely strigose and black petiole.

Chalcis sispes (Linnaeus, 1761)

Diagnosis – This species is easily distinguishable from *C. biguttata* by its red hind femur and ornamentation of the ventral margin.

Distribution – The species is widely distributed in Europe (Bouček, 1952) as well as Central Asia and Iran (Nikol'skaya, 1960).

Hosts – Ruschka (1920) quoting Giraud mentioned the stratiomyid fly *Stratiomys chamaeleon* (Linnaeus) as host for this species. Schremmer (1960) studied its oviposition behavior. The adults usually fly near ponds or rivers or standing waters, looking here for their hosts.

Cratocentrus tomentosus (Nikol'skaya, 1952)

Material examined – Syntypes ♀: "Lepidochalcis tomentosus Nik." / "Gourmouk, border between Persica (Iran), Afghanistan and Beloutchistan (Pakistan), 11.vi.1898, Zaroudni" [in Russian]. Syntypes ♂: "Lepidochalcis tomentosus Nik." / "Koucha-Liaroumba, Bampour, Iran (SE), 06-10.v.1901, Zaroudni" [in Russian].

Diagnosis – Female. Head, mesosoma, legs and antennae entirely reddish, scape darkened, mandibles black. Patches of dense, appressed and silvery setation masking the integument. Head without definite teeth on vertex, only low protrusions visible. F somewhat longer than wide, subequal. Pronotal collar with margins of punctures forming crests in middle; two blunt submedian teeth delimit a median furrow. Erect setae on mesonotum much longer than those of *Cratocentrus fastuosus* (Masi). Mesoscutum short, transverse, not longer than pronotal collar. Apical tooth of scutellum broad, triangular, rised upwards. Fore tibia

with acute apical spine. Mid tibia without apical spine, with subapical crests. T1 with sparse punctulation, interspaces much broader than points. Patches of appressed setation on T5 broadly separated. Basal part of syntergum, in front of cercal foveae, with lateral protrusions. Exerted part of ovipositor $\times 0.60$ mesosoma length.

Male. Syntergum deeply emarginate apically, between lateral projections.

This species would be keyed out using Steffan's key (1959b) as *C. fastuosus*. It differs from it in having great extension of reddish parts of body, low teeth on the vertex, submedian teeth on pronotal collar and characteristic morphology of the syntergum as described above.

Distribution – *Cratocentrus tomentosus* is known from India (Narendran, 1989) and Iran (Nikol'skaya, 1952).

Remarks – The species was originally described as *Lepidochalcis tomentosa* from northern part of Iran by Nikol'skaya (1952) and later transferred to *Cratocentrus* (Steffan, 1957).

Trigonura ninae (Nikol'skaya, 1952)

Trigonura sphenopterae Nikol'skaya, 1960 syn. n.

Material examined – Lectotype ♀: here designated, "*Urochalcis ninae* sp. n. type Nikol'skaya det." / "Répétek, 26.VII.1925, V. Gussakovski" [in Russian]. Syntypes 4 ♀♀: "*Trigonura sphenopterae* Nik." / "Bakanass-Ak-Kohl, Alma-Ata region, VIII.53, V. Parfentev" [in Russian].

Diagnosis – Female. Mandibles tridentate. Frons with dense puncturation, sparsely on gena. Base of antennal toruli on lower ocular line. Interantennal projection narrow, as high as toruli diameter. Inner margins of eyes subparallel. Scape reaching height of median ocellus. Occiput strongly sloping and concave, vertically striolate. Mesoscutum short, half as long as broad. Scutellum moderately convex. Propodeum without anterolateral tooth, with broad, depressed and rugose median area delimited by sublateral costulae; lateral surface of propodeum rugose areolate. Disk of hind femur smooth between the sparse piliferous points; setae suberect; ventral margin with 8-10 teeth. Fore wing with only microtrichiae, lacking fringe. T1-T6 smooth dorsally, T6 densely punctured. Cerci at mid length of syntergum. Relative measurements: head ×1.75 as broad as long, malar space ×0.75 breath of oral fossa and ×0.70 eye height. Eyes separated by 4/3 of their heights. OOL about as large as ocelli diameter. Pedicel+flagellum ×1.20-1.25 width of head. Mesosoma ×1.60 as long as wide.

Gaster ×1.50 length of mesosoma, syntergum 1/3 as long as gaster. Body reddish with black parts.

Distribution – The species is known from Caucasus and Central Asia (Nikol'skaya, 1952, 1960) and Iran (Haeselbarth, 1983).

Hosts – Trigonura ninae was reared from a buprestid beetle by M. Abai in Tehran, Iran.

Remarks – The two species were distinguished by Nikol'skaya through their differences in coloration. The body of lectotype of *U. ninae* is entirely reddish while the paralectotypes are more or less black. The type series of *T. sphenopterae* is extensively black, especially the mesosoma. The relative length of the funicular segments also is somewhat variable, with a relatively longer flagellum in the type series of *T. sphenopterae*. One of the paralectotypes of *U. ninae* is identical with this series.

Trigonura ruficaudis (Cameron, 1913)

Material examined – Iran: Yazd province, Ardakan - Zarch and Taft, ex: *Chrysobothris* sp. (Col.: Buprestidae), summer 1991, (Ahmadian), $7 \Leftrightarrow$ and $4 \Leftrightarrow$. HMIM (no label), $1 \Leftrightarrow$ and $1 \Leftrightarrow$.

Distribution – This species had been known only from India (Narendran, 1989) before being reported from Iran by Lotfalizadeh & Khalghani (2008).

Remarks – One female of this species was determined by G.J. Kerrich in 1966 as *Trigonura ruficaudis* which is labelled "A252, Mousl/IRQ, leg. H. Knopf", indicating that it was collected from Iraq.

II- Subfamily Dirhininae

Dirhinus himalayanus Westwood, 1836

Distribution – This species is known from Japan to Central Asia (Noyes, 2011) and Iran with some biological studies (Maadani & Ladonni, 1993; Dabiri *et al.*, 1998).

Hosts – Maadani & Ladonni (1993) and Dabiri *et al.* (1998) concluded that this species through its competitive biological characters such as high number of produced eggs, great searching ability and possible cold storage can be an ideal biocontrol agent against synanthropic flies, especially the house fly.

Dirhinus wohlfahrtiae Ferrière, 1935

Material examined – Iran: Ilam, vi.2004, ex: dipterous puparia, (B. Gharali), $2 \mathcal{Q}$.

Diagnosis – Head with broadly rounded horns. *Dirhinus wohlfahrtiae* and *D. himalayanus* are extremely close and further studies are needed – possibly the use of molecular data – to precisely separate the species.

Distribution – This species has been known from Egypt (El-Agoze, 1989), Saudi Arabia (Alahmed, 1999) and Sudan (Ferrière, 1935). This is the first record of this species from Iran.

Hosts – *Dirhinus wohlfahrtiae* was reared from puparia of Calliphoridae (Diptera) in Iran and is regarded as a biocontrol agent of house fly, *Musca domestica* L. Two other calyptrate Diptera, *Parasarcophaga aegyptiaca* (Salem) and *Wohlfahrtia nuba* (Wiedemann) have been reported as its hosts (Ferrière, 1935; El-Agoze, 1989; Alahmed, 1999).

III- Subfamily Haltichellinae

As in many Old World regions, Haltichellinae is the most diverse subfamily of Chalcididae in Iran (table 1).

Antrocephalus hypsopygiae Masi, 1928

Material examined – Iran: Mazandaran, Ghaemshahr, on *Oryza sativa*, 20.vii.2005, (S. Hesami), $1 \ \bigcirc$.

Distribution – *Antrocephalus hypsopygiae* was described from specimens from Turkmenistan (Masi, 1928) and has been reported from Central Asia to Cyprus (Bouček, 1956). GD examined additional specimens from southern France, Spain and Morocco. The genus *Antrocephalus* Kirby is recorded here for the first time from Iran.

Hosts – The examined specimen was collected in a rice field infested by rice stem borer, *Chilo suppressalis* Walker (Lep.: Crambidae) and green semilooper, *Naranga aenescens* Moore (Lepidoptera: Noctuidae) on the coastal area of Caspian Sea. It has was originally reared from Clover Hay worm, *Hypsopygia costalis* (Fabricius) (Lep.: Pyralidae) (Masi, 1928).

Belaspidia obscura Masi, 1916

Diagnosis – Body black. Malar space at least ×0.75 height of eye and as long as width of oral fossa. Mesonotum with moderately deep puncturation. Gaster at most 1.4 times as long as the thorax. T3 dorsally bare or very sparsely setose (Delvare *et al.*, 1999).

Distribution – Delvare *et al.* (1999) reported this species from western part of the Mediterranean Basin (from Europe to Middle East), and Tudor (1962) reported it from Iran.

Hosts – No definitive host is known for this species. The recorded hosts in the databases (Noyes, 2011) are the results of its misidentification with *Belaspidia nigra*.

Belaspidia nigra (Siebold, 1856)

Material examined – Iran: Azarbaijan-e Sharghi, Marand, Payam, 28.vi.2007, (H. Lotfalizadeh), 1 $\stackrel{?}{\circ}$. Esfahan, Esfahan, summer 1994, ex: Psychidae (Lep.), (Erfani), 6 $\stackrel{?}{\circ}$ and 3 $\stackrel{?}{\circ}$ $\stackrel{?}{\circ}$.

Diagnosis – Female. Body black. Malar space $\times 0.6$ height of eye, shorter than width of oral fossa ($\times 0.8$ in female and $\times 0.75$ in male). Mesonotum with moderately deep puncturation. Gaster $\times 1-1.15$ mesosoma length.

Distribution – *Belaspidia nigra* has been reported from Spain, Turkey and Azerbaijan (Delvare *et al.*, 1999) and also from Iran (Nikol'skaya, 1952).

Hosts – The species is a parasitoid of psychid moths, especially of the genus *Apterona* Millière.

Hockeria near bifasciata Walker, 1834

Material examined – Iran: Mazandaran, Ghaemshahr, 20.vii.2005, swept in rice field, (S. Hesami), $1 \ \bigcirc$.

Diagnosis – Its coloration is identical to *erdoesi* form Bouček (1952). Compared with *H. bifasciata*, it differs in having narrower hind femur, less projecting basal tooth. More specimens are needed for a certain identification.

Remarks – The above specimen was collected from rice field on the coastal area of Caspian Sea (in the same condition as for *A. hypsopygiae*). It is a parasitoid of moths (Lepidoptera).

Hockeria confusa Nikol'skaya, 1960

Material examined – Holotype ♀: "*Hockeria confusa* sp. n. type, det. Nikol'skaya" / by monotypy labelled "Iran, Gassmar, Tehran, VII.1955, Daftari" [in Russian].

Diagnosis – Species is close to *H. bifasciata*. Puncturation of vertex and mesonotum sparse, median lobe of mesoscutum interspaces double of points width, very small. Frenal

lobes strongly narrow, scutellum hardly emarginate. Anellus, F1 and legs reddish with fore and hind coxae darkened basally as well as hind tibia along ventral margin.

Distribution – Originally described from a single specimen and is known only from Iran (Nikol'skaya, 1960).

Hockeria unicolor Walker, 1834

Material examined – Lectotype \mathcal{E} : collected in France (BMNH type 5-293). Other material: Iran: Ilam, Shirvan - Chardavol, v.2003, Malaise trap in the field of safflower (*Carthamus oxycantha* M. B.), (B. Gharali), $1 \mathcal{L}$.

Distribution – The species is common and widely distributed in the Palaearctic region from Canary Islands (Báez & Askew, 1999) to Central Asia (Nikols'kaya, 1978).

Hosts – *Hockeria unicolor* is known as a pupal parasitoid of microlepidoptera of the families Tortricidae, Cosmopterigidae, Pyralidae (Bouček, 1952, 1977; Nikol'skaya, 1960; Blasco-Zumeta, 2000). The mention of a Curculionidae (Menozzi, 1947) results probably from a misidentification.

Kriechbaumerella gracilis (Nikol'skaya, 1952) comb. n.

Material examined − Holotype ♀: labelled "*Antrocephalus gracilis* sp. n. type Nikol'skaya det." / "Tavriz [Tabriz] Persica [Iran] 12 VI '14 P. Andrievski" [in Russian].

Diagnosis – Legs reddish except black coxae and brownish hind tarsus. Basal four antennomeres reddish, rest of antenna dark brown to black. The species is extremely close to *Kriechbaumerella mansues* (Nikol'skaya, 1952) but differs from in having the following characters: interspaces between points on mesonotum coriaceous and dull; scutellum with dorsal outline mostly straight, strongly sloping in front of frenal carina, relevant submedian lobes very short; apical margin of scutellum very shallowly emarginate; pale spot on fore wing larger, reaching apex of postmarginal vein; infuscation not reaching apex of wing; basal cell mostly bare except one row of setae behind submarginal vein; gaster slender, ×2.5 as long as broad.

Distribution – The species was described from Iran and is still known only from this country (Nikol'skaya, 1952).

Remarks – *Kriechbaumerella gracilis* was originally described as *Antrocephalus gracilis* by Nikol'skaya (1952) and later transferred to *Eucepsis* Steffan (Nikol'skaya, 1960). It

is here transferred to *Kriechbaumerella* Dalla Torre, the senior synonym of *Eucepsis* (Bouček, 1988).

Kriechbaumerella hofferi (Bouček, 1952)

Antrocephalus goliath Nikol'skaya, 1952 [synonymized by Bouček (1956: 239)]

Material examined – Lectotype ♀: here designated, a pinned female labelled "Antrocephalus goliath sp. n. type, det. Nikol'skaya" / "Kopet-Dag Komarovsk, Caspian Region, Turkmen. [istan], 24.VI.1928, V. Gussakovski" [in Russian].

Diagnosis – Eyes without distinctive pubescence. Antennae black. Flagellomeres relatively long, F1 ×2.5 as long as broad. Wing infuscated beyond parastigma with a pale spot below stigmal vein, inner margin delimited; infuscation still visible beyond venation. Hind femur black, contrasting with the relevant tibia which is reddish.

Distribution – The species is known from Europe to Central Asia. Nikol'skaya (1952) mentions it from Kazakhstan, Iran and Afghanistan nevertheless all the syntypes examined and labelled as such by her came from Turkmenistan. It is quoted from China as a parasitoid of *Caligula japonica* Moore (Lep.: Saturniidae) (Sheng, 1986).

Remarks – *Kriechbaumerella hofferi* was described in the genus *Eucepsis* (Bouček, 31/05/1952) and later transferred to *Kriechbaumerella* by Kalina (1989). *Antrocephalus goliath* was described from Turkmenistan by Nikol'skaya (30/06/1952) and later synonymized with *K. hofferi* by Bouček (1956).

Lasiochalcidia cincticornis (Walker, 1871)

Euchalcis rubripes Kieffer, 1899 [synonymized by Steffan (1966)]

Material examined – Syntype ♀: *Lasiochalcidia rubripes*, labelled "Bonifacio 1896 Ferton" / "*Lasiochalcidia cincticornis* (Walker), J.-R. Steffan det.", from the Ferton's collection itself (Steffan, 1951b). Other material: Iran: Sistan-Baluchestan, Bampur, 10.viii.1961, 1♀.

Diagnosis – This species belongs to the *dargelasii* species group which is mostly defined negatively: both apical spurs of hind tibia expanded as usual, apical serrulate lobe on hind femur not conspicuously expanded. Within this species group it can be distinguished by the male scape without tooth, anellus distinctly shorter than pedicel in female, sides of pronotum convex in dorsal view and strongly converging anteriorly, mesoscutum with irregular and moderately dense puncturation, scutellum slightly shorter than broad, shortly

truncate anteriorly and with evidently convex dorsal outline in lateral view, propodeum with the two lateral teeth appearing as broad and moderately protruding lobes. Otherwise the gaster is black, the legs are reddish with mid coxa and base of hind one reddish.

Distribution – The distribution mentioned in Noyes (2011) should be revised through re-examination of the collections as superficially similar species appears to be mixed by previous authors. *Lasiochalcidia cincticornis* itself might be restricted to the Mediterranean Basin, but has been recorded from Central Asia to Europe and North Africa.

Hosts – Bouček (1956) reported Lepidoptera pupae as hosts for *Lasiochalcidia* spp. but this is very doubtful as all known hosts for the genus are neuropteran Myrmeleontidae. Steffan (1958, 1959c, 1961, 1966) precisely studied the oviposition behaviour of several species and found that they tend to discover hidden hosts. The perception of the presence of a host occurs from the moving of sand subsequent to that of the antlion larva and the antennae bears special mechanoreceptor for that.

Remarks – The above female is considered here as a syntype because it bears the same information as quoted in the original description of *E. rubripes* by Kieffer (1899). Kieffer did not elaborate on the number of available specimens when he described the species, but most likely he had only a single female. *Euchalcis rubripes* has been variously interpreted since (Masi, 1916; Bouček, 1949, 1952, 1956; Steffan, 1951b, 1966). Steffan (1966) discussed those interpretations and concluded that *Halticella cincticornis* Walker and *E. rubripes* were synonyms. We confirm here Steffan's opinion. Firstly, the both species were described from Corsica. Walker (1871) mentioned in the original description of *H. cincticornis* "Antennae slender ... / ... first joint slender; second much longer than the third". The two species which were initially mixed but can easily be separated using the morphology of the male scape. That of *L. cincticornis* is unarmed while in the alternate [= *L. rubripes* sensu Masi, 1916 and Bouček, 1956] it bears a sharp ventral tooth. In the female of this second species anellus is hardly shorter than pedicel (×0.85) in contradiction with the type of *E. rubripes* and original description of *H. cincticornis*.

Lasiochalcidia indescripta Bouček, 1952 stat. rev.

Material examined – Paratype ♂: labelled "Lasiochalcidia indescripta Bouček Paratypus ♂ Z. Bouček 49" / "Szabotika Leg: A. Taubert" / "8.VII.1923 A. Taubert" / "Lasiochalcidia indescripta Bouček Bouček det. 1949". Other material: Iran: Gholestan, Gholestan National Park, Sulgerd, 1150 m, 20-21.vii.1996, (E. Ebrahimi), 1♀.

Diagnosis – *Lasiochalcidia indiscripta* can be recognized through the following characters: puncturation of mid lobe of mesoscutum progressively coarser from base to apex; scutellum with strongly convex dorsal outline in lateral view; lobes of frenal carina obviously raised upwards; metafemur entirely reddish; hind basitarsus short, ×3.5-3.8 as long as wide; gaster ×1.55-1.60 as long as wide; T2 slightly concave posteriorly.

Distribution – This new record expands the distribution of *L. indescripta* to Iran. The species was also known from central Europe (Bouček, 1952). GD examined specimens from southern France.

Hosts – Steffan (1966) studied the host recognition and oviposition behaviour. The hosts quoted by Steffan are Myrmeleontidae building traps such as *Myrmeleon insconspicuus* Ramur and *Neuroleon nostras* (Fourcroy).

Remarks – *Lasiochalcidia indiscripta* was synonymized with *L. guineensis* (Steffan) by Steffan (1953). The examination of the type, as well as other material, including both sexes, from southern France, showed that they are good species. Therefore, *L. indescripta* is here removed from synonymy.

Lasiochalcidia sparsibarbis Bouček, 1956

Material examined – Holotype ♀: labelled "Teheran Keredj" / "F. Brandt" / "Holotypus" / "Lasiochalcidia sparsibarbis Bouček sp. n. Det. Z. Bouček 1953".

Diagnosis – This species belongs to the same nominal species group which includes two strikingly close and apparently rare species from southern France and northeastern Spain. In this species group, the frons laterally to the antennal toruli shows relatively sparse setation which does not mask the integument. *Lasiochalcidia sparsibarbis* itself can be identified by its reddish hind leg, scape, pedicel and base of flagellum; a long malar space, slightly longer than width of oral fossa [×1.07 (45:42)]; lateral teeth of propodeum blunt and fairly protruding.

Distribution – This species is known only from the female holotype collected in Iran (Bouček, 1956).

Lasiochalcidia sp.

Material examined – Iran: Tehran, Varamin, Abardezh, 30.vi.1987, (M. Abai), 2 ♂♂.

Remarks – Two badly damaged males of *Lasiochalcidia* Masi run to *L. pubescens* (Klug, 1834) [= *L. igiliensis* (Masi)] according to Bouček's key (1952). Nevertheless several

characters led us to think that they belong to an undescribed species: (1) antenna with brown funicular segments (completely black in *L. pubescens*), (2) all funicular segments longer than wide (F5-F7 as long as wide in *L. pubescens*), (3) anellus less than ½ of length (wider than ½ of length in *L. pubescens*), (4) F1 ×1.5 as long as wide (×1.65 in *L. pubescens*), and (5) tibiae completely red (partly red in *L. pubescens*).

Neochalcis fertoni (Kieffer, 1899)

Material examined – Iran: Azarbaijan-e Sharghi, Marand, 1360 m, N 38° 25′ 28″ & E 45° 46′ 59″, 12.vii.2007, Malaise trap, (H. Lotfalizadeh), 1 ♂.

Diagnosis – Mesosoma entirely black, partly red or even nearly entirely red; gaster and legs black. Setation on head as usual, not especially long and erect. Interantennal projection high, strongly compressed, well visible in lateral view. Median areola of propodeum elongate, subelliptic. Epicnemial carina without medioventral projection. Apical spurs of hind tibia broadly separated at their insertion, by a distance larger than outer spur.

Distribution – The species is widely distributed in western and central Europe (Bouček, 1952), Mediterranean Basin (Bouček, 1956, 1970) and Central Asia (Nikol'skaya, 1978). The genus *Neochalcis* Kirby is newly reported from Iran.

Hosts – *Neochalcis fertoni* is a parasitoid of hymenopteran Aculeate nesting in twigs and dry stems (Bouček, 1970). The holotype of *N. fertoni* was reared from *Leptochilus* (*Neoleptochilus*) *regulus* (Saussure, 1855) [as *Odynerus gallicus* Saussure].

Neohybothorax hetera (Walker, 1834)

Material examined – Iran: Azarbaijan-e Sharghi, Marand, 1360 m, N 38° 25' 28" & E 45° 46' 59", 22-25.vii.2007, Malaise trap, (H. Lotfalizadeh), 1 $\, \bigcirc$. Azarbaijan-e Sharghi, Khosroshahr, 1346 m, N 37° 58' 28" & E 46° 02' 55", 10.x-6.xi.2007, Malaise trap, (H. Lotfalizadeh), 1 $\, \bigcirc$.

Distribution – The species has been recorded from Caucasus to Europe (Noyes, 2011). **Remarks** – *Neohybothorax hetera* attacks Ascalaphidae (Neuroptera) (Sellenschlo & Tröger, 1993) but our specimens were collected beside of dead wood highly infested by xylophagous beetles of the families Scolytidae and Bostrichidae. This species is recognisable by its rusty legs except coxae, distinct teeth of hind femur (especially in the last quarter) and uniformly rounded scutellum. In Iranian specimens collected in the northwester area, all legs are rusty (or reddish) as in Bouček's redescription (1952) with a greatly dark hind femur, reddish hind tibia with anterior longitudinal dark band; scape dark reddish and pedicel completely reddish.

Proconura caryobori (Hanna, 1934)

Material examined – Paratype \mathfrak{P} : labelled "London, ex: *Caryoborus pallidus* in senna pods from Sudan". Other material: Iran: Khuzestan, Ahvaz, summer 1998, ex: *Caryedon acaciae* (Gyllenhal), (Mozafari), $3 \mathfrak{P}$ and $1 \mathfrak{T}$.

Diagnosis – Tegulae and scape reddish. Flagellum slender with anellus ×2. Puncturation of pronotum and mesonotum sparse, interspaces larger than points on mid lobe of mesoscutum. Propodeum with posteroventral projections. Gaster lanceolate with T6 hardly sloping.

Distribution – Sudan from seeds of *Cassia*; India, Nepal (Narendran, 1986, 1989) and UK

Hosts – Caryedon pallidus (Olivier) (Col.: Chrysomelidae: Bruchinae) (on Cassia) was recorded as host (Hanna, 1934; Steffan, 1976; Bouček, 1988). Caryedon acaciae is a new host.

Remarks – This species was originally described from Sudan as *Euchalcidia caryobori* Hanna, but later transferred to the genus *Proconura* Dodd (Bouček, 1984). The biology and the effect of low temperatures on the fertility of this species were studied by Hanna (1934). Hence, *P. caryobori* is one of the few Chalcididae for which some detailed studies on the biology was carried out.

Proconura incongruens (Masi, 1932)

Material examined – Iran: Kerman, Jiroft, 630 m, N 28° 28′ 57″ & E 57° 52′ 08″, 03.xi.2009, (Rajabi), 1 ♂.

Diagnosis – The male of this species can be easily recognized through deeply excavated scape on ventral side (Steffan, 1976: fig. 14, p. 59).

Distribution – The species was previously known only from Lybia. This new record hence greatly expands its distribution area.

Proconura nigripes (Fonscolombe, 1832)

Material examined – Iran: Azarbaijan-e Sharghi, Khosroshahr, 1346 m, N 37° 58′ 28″ & E 46° 02′ 55″, 10.x-6.xi.2007, Malaise trap, (H. Lotfalizadeh), 2 ♂♂.

Diagnosis – Body and tegula dark. Malar space long. Flagellum slender, with anellus and F1 much longer than wide. Propodeum with lateral protrusions and posteroventral tooth; median areola long. Gaster oval, acute with T6 and T7 visible from above (Steffan, 1976). The male cannot be separated from males of *P. caryobori*.

Distribution – *Proconura nigripes* is widely distributed in the Palaearctic, having been reported from the Canary Islands (Baez & Askew, 1999), whole Europe (Steffan, 1951b; Bouček, 1952), North Africa (Masi, 1929) and Central Asia (Nikol'skaya, 1978).

Hosts – Davatchi reared this species from *Schneidereria pistaciicola* (Danilewski, 1955) (Lep.: Gelechiidae) in Iran (Davatchi, 1958).

Proconura persica Delvare, 2011

Material examined − Holotype ♀: Iran: Isfahan, Kashan, viii.2009, ex: *Apomyelois ceratoniae* (Zeller) (Lep.: Pyralidae) on *Ceratonia siliqua* (L. Talaee).

Distribution – The species was recently described (Delvare *et al.*, 2011) and is known only from Iran as a parasitoid of carob moth.

Psilochalcis ceratoniae Delvare, 2011

Distribution – The species was recently described (Delvare *et al.*, 2011) and is known only from Iran as a parasitoid of carob moth.

Psilochalcis ligustica (Masi, 1929)

Material examined – Iran: Ilam, Shirvan - Chardavol, Malaise trap, vi.2003, (B. Gharali), 1 ♀. Qazvin, Juladak, N 36° 21' 53" & E 50° 32' 11", 14.vi.2007, (B. Gharali), 1 ♂.

Diagnosis – This species belongs to the same nominal species group, in which the mandibles are strongly incised on their ventral margin, suggesting an additional tooth.

Distribution – *Psilochalcis ligustica* was reported from Iran as *Invreia ligustica* Masi (Bouček, 1952, 1956), and from Kurdistan and Shaklawa. It is widely distributed in Palaearctic and India (Narendran, 1989).

Psilochalcis subaenea (Masi, 1929)

Material examined – Iran: Khuzestan, Ahvaz, summer 1998, ex: *Caryedon acaciae* (Gyllenhal), (Mozafari), $4 \Im$.

Diagnosis – Head with a short triangular frontal lobe below the antennal toruli, hind tibia with an additional external carina, coxae and femora black [the usual ventral outer carina is visible near base of the hind tibia while the additional carina can be seen near the apex].

Distribution – This species is distributed in the Mediterranean region from Spain to Kazakhstan (Masi, 1929; Bouček, 1952, 1956; Nikols'kaya, 1960) and is a new record for the Iranian fauna.

Hosts – The specimens were reared from *C. acaciae* in southwestern Iran.

Remarks – This species was originally described as *Invreia subaenea* and designated as the type species of the new genus *Invreia* by Masi (1929). It was later synonymized with *Psilochalcis* Kieffer by Bouček (1992). Biology of this species is unknown.

Psilochalcis subjecta (Nikol'skaya, 1960) comb. n.

Material examined – Holotype ♀: labelled "Khorasa 3-4.VIII.1901 N. Zaroudni" [in Russian] / "Invreia subjecta sp. n. ♀ Nikols'kaya det." / "Holotypus ♀".

Diagnosis – Body and fore coxa black; tegula, rest of legs and antenna reddish. Gaster reddish-brown. Head long ×2 with long temples merging to occiput without evident angulation. Carina delimiting clypeus dorsally not extended to triangular flange (it is extended in *P. subaenea*). Malar space slightly more than half as long as height of eyes. Antennal scrobes not reaching median ocellus. Pedicel elongate ×4. Flagellum relatively long and slender: anellus ×2, F1 ×1.6. Propodeum with posterolateral angles broadly blunt, lateral and sublateral carinae subparallel, submedian hardly curved. Hind tibia with additional outer carina. Gaster acute, dorsal outline moderately sloping posteriorly.

Distribution – *Psilochalcis subjecta* is known only from Iran (Nikol'skaya, 1960).

Remarks – This is probably the female of *Psilochalcis miranda* (Nikol'skaya, 1960) which was described from the male; a paratype of which was examined by GD.

Psilochalcis zarudnyi (Nikol'skaya, 1960) comb. n.

Material examined – Paratype \cite{Q} : labelled "Koucha-Liaroumba, Bampour Youv Persia Zaroudni 6-10 V 01" [in Russian] / "*Peltochalcidia zarudnyi* sp. n. \cite{Q} Nikols'kaya det." / "Paratypus \cite{Q} ".

Diagnosis – Legs, antennae and tegulae completely reddish. Mandibles with a distinct furcate carina on the disk, without subapical swelling on lower tooth. Frontal lobe broader than long, with a few thick basal setae. Head ×2 in dorsal view with temples strongly converging. Posterior margin of propodeum forming obtuse but sharp posterolateral angles. Gaster ovoid, with T6 strongly sloping, almost vertical.

Distribution – Nikol'skaya (1960) described the species based on material collected from Iran. It was apparently not collected again.

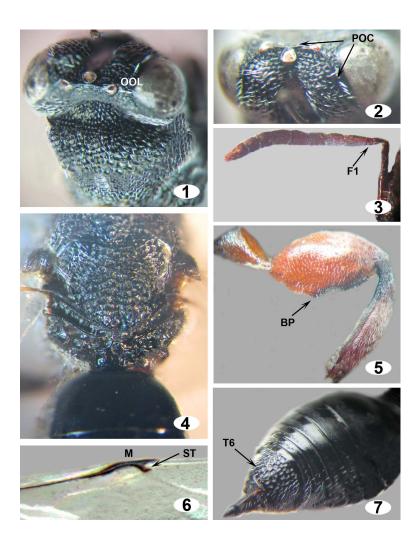
Remarks – Steffan (1951c) synonymized the species *Peltochalcidia ferrierei* Steffan with *Peltochalcidia benoisti* Steffan. The former species was described from Spain while *P. benoisti* was from north Africa. The examination of the holotypes of the two species, together with additional specimens, showed that the specimens distributed on each side of the Mediterranean Sea are morphologically different. The species remain valid while *Peltochalcidia oranensis* Bouček, 1952 is found to be a junior synonym of *P. benoisti* syn. n. The species *Psilochalcis ferrierei* (Steffan) is revalidated here, stat. n. and comb. n.

Psilochalcis zarudnyi belongs to the *benoisti* species group, in which a projecting frontal lobe overlaps the clypeus and the base of the mandibles. This species is very close to them, especially the frontal lobe, which has a similar habitus but is slightly longer.

Tanycoryphus tibialis (Nikol'skaya, 1960) comb. n. (Figs 1-7)

Material examined – Paratype $\$: labelled "Ou. Kondara 1100 m, D. Varzoba, Tadj. [ikistan] Gussakovskyi" [in Russian] / "*Varzobia tibialis* sp. n. $\$ Nikols'kaya det." / "Paratypus $\$ ". Other material: Iran: Tehran, Evin, 22.vi.2005, ex: Buprestidae (Coleoptera), on pine, (M. Abai), $\$ 4 $\$ 9 $\$ 0.

Diagnosis – Body and antenna black. Tegula, T1, T2, T3 laterally and syntergum reddish on examined paratype. Hind leg mostly reddish (fig. 5), fore and mid legs partly reddish on. Pre- and postorbital carinae and transverse blunt carina between lateral ocelli visible (fig. 2, POC). All F longer than wide, F1 ×2 as long as pedicel (fig. 3, F1). Walls of punctures forming transverse crests only on mid lobe of mesoscutum. Dorsellum with a large median areola. Fore wing bearing only microtrichiae, without fringe. M ×3 ST (fig. 6, M and ST). Hind femur with blunt basal tooth and one serrulate apical lobe. Additional outer carina of hind tibia complete although evanescent at base. Gaster ×1.85. T6 densely punctured (fig. 7, T6). Postcercale ×1.5 distance between cerci.



Figures 1-7. *Tanycoryphus tibialis* (Nikol'skaya). 1. Head and pronotum in fronto-dosral view; 2. head in dorsal view; 3. female antenna in lateral view; 4. mesonotum, propodeum and first gasteral tergite in dorsal view; 5. hind leg in lateral view; 6. forewing venation; 7. female gaster in dorsal view. BP = basal projecting of hind femur; F1 = first funicular segment; M = marginal vein; OOL = ocellar-ocular distance; POC = pre-orbital carina; ST = stigmal vein; T6 = sixth gastral tergite.

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Chalcidinae Brachymerini Brachymeria Wessbood B eccuration Chlore Appropriate (Lep.: Phatilate) B. control (Brachymeria Wessbood) B. control (Brachymeria) B. control (Brachymeria Wessbood) B. control (Brachymeria Wessbood) B. control (Brachymeria) B. control (Brachymeria Wessbood) B. control (Brachymeria)	Chalcidinae	canne	Genera	Species	Host
B. eccurinata Galan * B. eccurinata Galan * B. femorata (Panzer) B. fasas (Walker) B. minute (Limaeus) B. minute (Limaeus) B. minute (Limaeus) B. nighata (Ferster) * B. minute (Limaeus) B. nighata (Ferster) * B. nighata (Ferster)		Brachymerini	Brachymeria Westwood	B. ceratoniae Delvare	Apomyelois ceratoniae (Lep.: Pyralidae)
B. Jemorata (Panzer) B. Jemorata (Panzer) B. Jemorata (Passer) * B. minuta (Linnaeus) B. pervica (Masi) B. nitigataer (Sister) * B. nitigataer Spinela C. sispes (Linnaeus) Cratocentrial Cratocentr				B. excarinata Gahan *	Plutella xylostella (Lep.: Plutellidae)
Chalcidini Chalcis Fabricius B. Itsus (Walker)* B. minuat Chimaeus) B. persica (Masi) B. persica (Masi) B. persica (Masi) B. prinquence (Yasis comb. n. B. rufigatiosa (Forster)* B. prinquence (Nasi) comb. n. B. rufigatiosa (Forster)* B. prinquence (Spoura Sichel C. foguata Spinda C. sispese (Limaeus) Dirthimi Dirthimus Dalman D. himaloguana Westwood D. wohlfabriae Ferrière* Haltichellini Antrocephalus Kirby B. obscura Masi Belaspidia Masi B. nigat (Sibold) Hockeria Walker H. confusa Nikol'skaya (Masi shaga (Special Walker)* Lasiochalcidia Masi L. indescripta (Bouček) Lasiochalcidia Masi P. caryobori (Hama) * Proconura Dodd P. persica Delvare Tanycoryphus Cameron T. tibidis Nikol'skay comb. n. Persica Delvare Tanycoryphus Cameron T. tibidis Nikol'skay comb. n. Persica Delvare Tanycoryphus Cameron T. tibidis Nikol'skay comb. n. Persica (Masi) P. subcasa (Masi P. subcasa (Masi)) P. subcasa (Masi P. subcasa (Ma				B. femorata (Panzer)	Heliothis sp. (Lep.: Noctuidae)
Chalcidini Chalcis Fabricius B minuta (Linnaeus) R obnasara (Forster)* B persica (Masi) B ruglasster (Masi) comb. n. C rigitatus Sinda C sisper (Linnaeus) C rigitatus (Sinda) C rigitatus (Sinda) C rigitatus (Saraya) syn. n. T ruffcaudis (Cameron) * D rigitatis (Cameron) * D ruglastrate Ferriere * Hartichellini Antrocephalus Kirby Hockeria Walker A. hypsopygiae Masi * Belaspidia Masi B. obserut Masi * B rigitat (Siebold) Hockeria Walker H. unicolor Walker * H. unicolor Walk				B. lasus (Walker) *	
Chalcidini Chalcis Fabricius B. ohusana (Förster) * B. ohusana (Förster) * B. rufigaster (Nas) comb. n. B. rufigaster (Nas) B. rufigaster (Nas) B. rufigaster (Nas) Cratocentrini Cratocentrus Cameron C. sispes (Linnaeus) C. sispes (Linnaeus) C. sispes (Linnaeus) C. sispes (Linnaeus) C. prigutata Spincia Cratocentrini Cratocentrus Dalman D. ruficaudis (Cameron) * Relaxpidia Masi B. dokeara Masi * B. dokeara Masi B. dokeara Masi * B. ruficaudis (Siebold) Hockeria Walker H. confusor Nikol skaya H. unicolor Walker * R. gracifis (Nikolskaya) comb. n. K. pagrailis (Nikolskaya) comb. n. C. ruticeronis (Walkey) D. ruticidia sp. * L. raparsibarbis Boucek Lasiochalcidia sp. * L. sparsibarbis Boucek Lasiochalcidia sp. * Neochalcis Kirby N. Jertoni (Kiefer) * Procomura Dodd P. persica Delvare Tanycoryphus Cameron P. ligustica (Masi) P. ribidis (Nasi) P. ruficalaria Delvare P. sincara (Nasi) P. ruficalaria Delvare P. sincara (Nasi) P. ruficalaria Cameron P. ligustica (Masi) P. ruficalaria Comb. n. * P. subcara (Nasi) P. ruficalaria Delvare P. subcara (Nasi) P. ruficalaria Comb. n. * P. subcara (Nasi)				B. minuta (Linnaeus)	Lymanperia dispar (Lep.: Lymanteridae)
Chalcidini Chalcis Fabricius B. previca (Masi) B. rufigaster (Masi) comb. n. B. rufigaster (Masi) emb. n. B. rufigaster (Masi) emb. n. B. rufigulosa (Förster) * B. rufigulosa (Förster) * B. rufigulosa (Förster) * B. rufigulosa (Förster) * C. tomentosus (Nikolskaya) Phasgonophorini Trigonura Sichel T. rilnae (Nikolskaya) Dirhimin Dirhimus Dalman D. himalquanus Westwood D. wohlfahritae Ferriere * Hockeria Walker H. confusa Nikol'skaya emb. n. Belaspidia Masi B. doscura Masi * B. doscura Walker H. confusa Nikol'skaya comb. n. C. tenticorenis (Walker) * C. tenticorenis (Walker)				B. obtusata (Förster) *	
B. rufguester (Masi) comb. n. B. rufguester (Masi) comb. n. B. rigulosca (Ferster) * B. hibialis (Walker) Cratocentrini Cratocentrus Cameron C. opentrus Spinda Cratocentrus Sichel T. rune (Nikol Saxay) syn. n. T. rinne (Nikol Saxay) syn. n. T. rufacudis (Cameron) * Dirhinini Dirhinus Dalman D. himalayanus Westwood Dirhinini Dirhinus Dalman D. himalayanus Westwood Dirhinini Dirhinus Dalman D. himalayanus Westwood Belaspidia Masi B. obscura Masi * Belaspidia Masi B. obscura Masi * Rockeria Walker H. confines Nikol Saya * H. unicolor Walker * R. paresific (Nikol Saya) comb. n. Dalla Torre Dalla Torre A. hypocygue Masi * R. gracilis (Nikol Saya) comb. n. K. hoffer (Bouček) Lasiochalcida Masi L. cincticornis (Walker) * L. indexcripta (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcida syn. A. Regroulidis sp. * Neorhbothorax Nikol Skaya N. hereat (Walker) Proconura Dodd Proconura Dodd				B. persica (Masi)	
Chalcidini Chalcis Fabricius C. biguitata Spindales (Förster) * B. vitripennis (Förster) * C. siopes (Linnaeus) Cratocentrini Cratocentrus Cameron C. tomentasse (Nikolskaya) Phasgonophorni Trigonura Sichel T. rigicaudis (Cameron) * Dirhinini Dirhinus Dalman D. himalquanus Westwood D. violightrituse Ferriere * Haltichellini Antrocephalus Kirby A. hipsopygiae Masi * Belaspidia Masi B. o'secura Masi Belaspidia Masi Belaspidia Masi B. A. hipsopygiae Masi * Riccheria Walker H. confusa Nikol skaya H. micolor Walker * Kriechbaumerella K. gracilis (Nikolskaya) comb. n. C. norticennis (Walkeys) Lastochalcidia Masi L. cinetteennis (Walkeys) L. sparsibarbis Boucek Lastochalcis Kirby Neothalcis Kirby Neotholovat Nikolskaya N. herea (Walker) Proconura Dodd Proconura Dodd Priconinia Persica Delvare Tanyconyphus Cameron T. ibidis Sikaya comb. n. * P. infinity Consortione Delvare P. ingustica (Masi) P. subcane (Masi) P. su				B. rufigaster (Masi) comb. n.	
Chalcidini Chalcis Fabricius C, biguttata Spinela Cratocentrini Cratocentra Cameron C, sispes (Linnaeus) Phasgonophorini Trigonura Sichel T, ninae (Nikolskaya) Phasgonophorini Trigonura Sichel T, ninae (Nikolskaya) syn. n. T rifficaudis (Cameron) * T riffication * T rifficati				B. rugulosa (Förster) *	Apomyelois ceratoniae (Lep.: Pyralidae)
Chalcidini Chalcis Fabricius C bigutata chinaeus) Cratocentrini Cratocentrus Cameron C comentosus (Nikol Skaya) Phasgonophorini Trigonura Sichel T rupteculis (Cameron) * Dirhinini Dirhinus Dalman D himaloyamus Westwood Dirhinini Dirhinus Dalman D himaloyamus Westwood Dirhinini Dirhinus Dalman D himaloyamus Westwood Haltichellini Antrocephalus Kirby A hypeopygiae Masi * Belaspidia Masi B obserura Masi B obserura Masi * Relaspidia Masi B obserura Masi B nigar (Siebold) Hockeria Walker H conflusa Nikol Skaya H unicolor Walker * R Kriechbaumerella K gracilis (Nikol Skaya) comb. n. Dalla Torre Dalla Torre Lasiochalcidia Masi L. cincticornis (Walker) * Lasiochalcidia Masi L. cincticornis (Walker) * Lasiochalcidia Masi L. cincticornis (Roleige) * Neorhbothorax Nikol Skaya A reversibarbis Boucek Lasiochalcidia sp. * Lypusibarbis Boucek Lasiochalcidia sp				B. tibialis (Walker)	Euproctis chrysorrhoea, Euproctis melania, Leucoma wiltshhirei, Ocneria sp.,
Chalcidini Chalcis Fabricius C. higuttana Spinola Cratocentrini Cratocentrua Cameron C. tomentosus (Nikolskaya) Phasgonophorinii Trigonura Sichel T. ninae (Nikolskaya) sv. n. Dirhinini Dirhinus Dalman D. ninaediyamus (Nakolskaya) sv. n. T. ruficaudis (Cameron) * Dirhinini Dirhinus Dalman D. himadiyamus (Wastwood Haltichellini Antrocephalus Kirby A. hypsopyajaa Masi * Belaspidia Masi Belaspidia Masi Belaspidia Masi Belaspidia Masi Belaspidia Masi A. hypsopyajaa Masi * Ricechaumerella B. nigra (Sichold) Hockeria Walker H. conjusa Nikolskaya tunicolor Walker A. kyawaiis (Nikolskaya) comb. n. Dalla Torre Dalla Torre C. indexicolor Walker A. hypsopyajaa (Bouček) stat. rev. * Lasiochalcidia Masi L. cincticornis (Walker) * Lasiochalcidia Masi L. cincticornis (Walker) Proconura Dodd B. ryawisa Delvare Tanyconphus Cameron P. rayawisa Conbert P. persica Delvare Tanyconphus Cameron T. tibidis Nikolskaya comb. n. * Pripalia Proconura Dodd P. persica Delvare P. pragica (Masi) P. ribidis Nikolskaya comb. n. p. r					Parocneria terebynthina and Euproctis melania (Lep.: Lymanteridae), Zygaena
Chalcidini Chalcis Fabricius C. higurtaus Spiniolas Cratocentrini Cratocentrus Cameron C. sispes (Linnaeus) Cratocentrini Trigonura Sichel T. rintee (Nikolskaya) syn. n. Dirhinini Dirhinus Dalman D. himadionurae (Nikolskaya) syn. n. T. rigicaudis (Cameron) * Dirhinini Dirhinus Dalman D. himadionurae Wastwood D. wohligheritae Feriree * Haltichellini Antrocephalus Kirby A. hipsopygiae Masi * Belaspidia Masi B. obscura Masi * B. obscura Nikol skaya H. unicolor Walker * H. confusor Nikol skaya onnb. n. C. indicteorius (Valkay) * C. indicteorius (Walker) * L. sparsiburbis Bouček Lasiochalcidia Masi L. sparsiburbis Bouček Lasiochalcidia sp. * Neochalcis Kirby N. fertoni (Kriefet) * Proconura Dodd P. indiapies (Fonsolombe) P. persica Delvare Tanycoryphus Cameron P. ligansica (Masi) P. nigipies (Fonsolombe) P. persica Delvare P. sindende Oklasi) P. subcane (Masi) P. subcane (Masi)					napernauen (Lep.: Lygaenidae)
Cratocentrini Cratocentra Cameron C. togutuda spindal C. sispese (Linnaeus) Cratocentrini Trigonura Sichel T. intue (Rikolskaya) syu. n. Dirthinini Dirthinus Dalman D. himalcyanus Westwood D. wolifiantiare Ferrière* Haltichellini Antrocephalus Kirby Belaspida Masi Belaspidia Masi Belaspidia Masi B. obscura Masi B. nigat (Siebold) Hockeria Walker H. confusa Nikol'skaya H. unicolor Walker H. confusa Nikol'skaya H. unicolor Walker K. gracifis (Nikolskaya) comb. n. Dalla Torre K. forgeria Nanovaka (Nikolskaya) comb. n. Cratoconnis (Nikolskaya) comb. n. K. hofferi (Bouček) Lasiochalcidia Masi L. indescripta (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcidia Masi L. indescripta (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcidia Masi Proconura Dodd Proconura Dodd P. caryobori (Hama) * Proconura Dodd P. persica Delvare Tanycoryphus Cameron T. tibidis Nikol'skaya comb. n. * P. persica Delvare P. persica (Masi) P. sinkenger (Masi) P. subcaren (Masi) P. subcaren (Masi) P. subcaren (Masi)				B. vitripennis (Förster) *	
Cratocentrini Cratocentra Cameron C. tomentosus (Nikolskaya) Phasgonophorini Trigonura Sichel T. iniae (Nikolskaya) syn. n. Dirhimus Dalman D. himadayanus Westwood D. wohlfahritae Ferrière * Haltichellini Antrocephalus Kirby A. hippoopyigue Masi * Belaspidia Masi B. obscura Masi B. nigar (Siebold) Hockeria Walker H. confliae Nikolskaya B. nigar (Siebold) Hockeria Walker H. confliae Nikolskaya H. unicolor Walker * K. gracilis (Nikolskaya) comb. n. K. gracilis (Nikolskaya) comb. n. K. hafferi (Boucèk) Lasiochalcidia Masi L. cincticornis (Walker) L. sparsibarbis Boucèk Lasiochalcidia Stirby Neothybothorax Nikolskaya N. hetera (Walker) Proconura Dodd Proconura Dodd Pression (Palma) * Pricongruens (Masi) Pricongruens (Mas		Chalcidini	Chaicis Fabricius	C. orgunara spinola	
Cratocentrum Cratocentrus Cameron C. tomentosus (Nkolskaya) Phasgonophorini Trigonura Sichel T. rintnee (Nikolskaya) syn. n. Dirhinini Dirhinus Dalman D. himadiquania (Cameron)* Dirhinini Dirhinus Dalman D. himadiquania Westwood D. wohligheritae Feriree* Haltichellini Antrocephalus Kirby A. hipsoppigiae Masi * Belaspidia Masi B. obscura Masi * B. obscura Walker * H. confusor Nikol skaya H. unicolor Walker * H. confusor Nikol skaya omb. n. E. k. farcific (Nikolskaya) comb. n. E. hinderscipul (Bouček) * E. spansibarbis Bouček *			1	C. sispes (Linnaeus)	
Phasgonophorini Trigonura Sichel Trinnea (Noishsaya) syn. n. Dirhinini Dirhinus Dalman D. himdalyama Westwood Dirhinini Durhinus Dalman D. himdalyama Westwood Haltichellini Antrocephalus Kirby Belaspidia Masi Belaspidia Masi B. hypeopygiae Masi * Belaspidia Masi B. hypeopygiae Masi * Belaspidia Masi B. obserna Masi B. nigar (Siebold) Hockeria Walker H. conflias Nikol Skaya H. unicolor Walker * Kriechbaumerella K. gracilis (Nikol Skaya en H. unicolor Walker * Lasiochalcidia Masi L. cincticornis (Walker) * Lasiochalcidia Masi L. cincticornis (Walker) * Lasiochalcis Kirby K. hofferi (Bouček stat. rev. * L. sparsibarbis Bouček Lasiochalcidia sp. * Neorhoboliorar Nikol Skaya N. hetera (Walker) Proconura Dodd P. caryobori (Hama) * P. persica Delvare Tanycorphus Cameron T. tibidis Nikol Skaya comb. n. * P. persica Delvare P. persica Delvare P. persica Oblare P. persica		Cratocentrini	Cratocentrus Cameron	C. tomentosus (Nikol'skaya)	
Dirthinini Dirthinus Dalman T. ruficandis (Cameron) * Dirthinini Dirthinus Dalman D. himaloyamus Westwood D. woldighrituse Ferriere * Antrocephalus Kirby A. hipsopygiae Masi * Belaxpidia Masi B. obserand Masi B. obserand Masi B. nigra (Siebold) Hockeria Walker H. confuso Nikol Skaya H. unicolor Walker * H. confuso Nikol Skaya H. unicolor Walker * H. confuso Nikol Skaya H. indexcripta (Boucek) L. sparsibarbis Boucek Lasiochalcidia Masi L. indexcripta (Boucek) L. sparsibarbis Boucek Lasiochalcidia sp. * Neochalcis Kirby N. Jeroni (Kieffer) * H. popropori (Hama) * P. persica Delvare Tanyconyphus Cameron T. indials Sinos (Masia) P. incongruens (Masi) P. persica Delvare Tanyconyphus Cameron P. ligustica (Masi) P. subcaned (Masia)		Phasgonophorini	Trigonura Sichel	T. ninae (Nikol'skaya) syn. n.	Buprestidae (Col.)
Dirhinini Drihinus Dalman D. himalquanus Westwood Dirhinini Antrocephalus Kirby A. hypoopygae Nasi* Belaspidia Masi B. obscura Masi * B. obscura Masi * B. obscura Masi * B. obscura Masi * B. nigar (Siebolt) Hockeria Walker H. confusa Nikol'skaya H. unicolow Walker * H. unicolow Wal				T. ruficaudis (Cameron) *	Chrysobothris sp. (Col.: Buprestidae)
Haltichellini Antrocephalus Kirby D. wohlfahrtiae Ferrière * Belaspidia Masi B. obseura Masi * B. nigra (Siebold) Hockeria Walker H. conflias Nikol'skaya H. unicolor Walker * K. gracifis (Nikol'skaya) comb. n. Dalla Torre K. gracifis (Nikol'skaya) comb. n. K. hoffer! (Boucèk) Lasiochalcidia Masi L. cinctirornis (Walker) * L. sparsibarbis Boucèk J. stat. rev. * Neochalcis Kirby N. fertoni (Kiefter) * Neochalcis Kirby N. fertoni (Kiefter) * Proconura Dodd P. caryobori (Hama) * Proconura Dodd P. caryobori (Hama) * Proconura Dodd P. raypes (Fonscolombe) P. persica Delvare Tanycoryphus Cameron T. tibialis Nikol'skaya comb. n. * P. sinbarate (Masi) P. subcarate (Masi)	irhininae	Dirhinini	Dirhinus Dalman	D. himalayanus Westwood	Musca domestica (Dip.: Muscidae)
Haltichellini Antrocephalus Kirby A. hypsopygiae Masi * Belaspidia Masi B. dokeura Masi Belaspidia Masi B. dokeura Masi B. nigra (Siebold) Hockeria Walker H. confiasa Nikol skaya H. unicolor Walker * H. confiasa Nikol skaya H. unicolor Walker * H. confiasa Nikol skaya H. unicolor Walker * L. sparsibarbis Boucek L. sparsibarbis Boucek Lastochalcis Kirby Neohalcis Kirby N. Jerton (Kieffer) * Neoholorax Nikol skaya N. herea (Walker) Proconura Dodd P. persica Delvare Tanycoryphus Cameron P. infiaits Nikol skaya comb. n.* Hybothoracini Psilochalcis Kieffer P. crantomica Delvare Psilochalcis Kieffer P. crantomica Delvare P. ligastica (Masi) P. sukheane (Masi)				D. wohlfahrtiae Ferrière *	Calliphoridae (Dip.)
Belaspidia Masi B. obscura Masi B. nigas (Siebold) Hockeria Walker H. orginas Nikol Skaya H. unicolow Walker* K. gracilis (Nikol Skaya) Lasiochalcida Masi L. cincticornis (Walker)* Lasiochalcida Masi L. cincticornis (Walker)* L. indescripta (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcida sp. * Neorhbothorax Nikol Skaya N. pervai (Kaffer) * Neorhbothorax Nikol Skaya N. pervai (Kaffer) * N. pervai (Kaffer) * N. pervai (Kaffer) * N. pervai (Kaffer) * Proconura Dodd P. persica Delvare Tanycorphus Cameron T. tibidis Nikol Skaya comb. n.* Hybothoracini Psilochalcis Kieffer P. persica Delvare P. persica Delvare P. persica Obelvare P. persica Obelvare P. persica Obelvare P. persica (Masi) P. subacute (Masi) P. subacute (Masi)	altichellinae	Haltichellini	Antrocephalus Kirby	A. hvnsonvoiae Masi *	Chilo supressalis (Len.: Crambidae) and Naranga agnescens (Len.: Noctuidae)?
Hockeria Walker Hockeria mear bifasciata Walker Hockeria near bifasciata Walker Hockeria near bifasciata Walker Horderia near bifasciata Walker Horder Walker Horder Walker Horder Bouckel) Lasiochalcida Masi Lindecornia (Walker) * Lindecornia (Walker) * Lindecornia (Walker) * Lindecornia (Boucke) stat. rev. * Lindecorpia (Boucke) stat. rev. * Lindecorpia (Boucke) stat. rev. * Lindecorpia (Boucke) stat. rev. * Lindecornia (Boucke) stat. rev. * Lindecorpia (Boucke) stat. rev. rev. rev. rev. rev. rev. rev. rev			Belasnidia Masi	B. obscura Masi	
Hockeria Walker In graft of social Walker Hockeria near bifasciata Walker Hockeria near bifasciata Walker Hockeria Nikol'skaya Humicolow Walker* Kriecirbaumerella K. gracilis (Nikol'skaya) comb. n. K. hofferi (Bouček) Lasiochalcidia Masi L. indescripta (Bouček) L. indescripta (Bouček) L. sparsibarbis Bouček L. sparsibarbis L. sparsib			Town and done	R mirra (Siahold)	Derrohidae (Lan)
Hockerta Walker Hockerta uber plascatta Walker H. conflist Nikol'skaya H. unicolor Walker K. gracific Nikol'skaya) comb. n. Dalla Torre K. gracific Nikol'skaya) comb. n. K. lofferi (Bouček) L. sparsibratis (Walker) L. sparsibratis Bouček Lasiochalcidia sp. K. parsibratis Souch R. parsibratis (Masi) R. parsibratica (Masi) R. subuenen (Masi) R. parsibratica (Masi) R. subuenen (b. nigra (Siebold)	Fsychidae (Lep.)
H. conflusa Nikol's skaya H. unicolor Walker * Kriechhaumerella K. gracilis (Nikol'skaya) comb. n. Dalla Torre K. hofferi Boucek) Lasiochalcidia Masi L. cincticornis (Walker) * L. sparsibarbis Boucek Lasiochalcidia sp. * L. sparsibarbis Boucek Lasiochalcidia sp. * Neotybothorax Nikol'skaya N. hereat (Walker) Procomura Dodd P. carpobori (Hanna) * P. carpobori (Hanna) * P. carpobori (Hanna) * P. meorgraens (Masis) P. mersica Delvare Tanycoryphus Cameron P. persica Delvare P. persica (Masis) P. sulvaenet (Masis)			Hockeria Walker	Hockeria near bifasciata Walker	Chilo supressalis (Lep.: Crambidae) and Naranga aenescens (Lep.: Noctuidae)?
Kriechbaumerella K. gracilis (Nikol'skaya) comb. n. Dalla Torre Lasiochalcidia Masi L. cineticornis (Walker)* Lindescriptu (Bouček) stat. rev. * L. indescriptus (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcidia sp. * Lasiochalcidia sp. * Neohybothorax Nikol'skaya N. Jeroni (Koeffer) * Proconura Dodd P. in congruense (Mais) Proconura Dodd P. in congruense (Mais) Procoluse (Mais) P. persica Delvare Psilochalcis Kieffer P. persica Delvare Psilochalcis Kieffer P. persica Delvare Psilochalcis Kieffer P. eventomica Delvare				H. confusa Nikol'skaya	
Kriechbaumerella K. gracilis (Nikol'skaya) comb. n. Dalla Torre K. hoffer (Bouček) Lasiochalcida Masi L. indexcripta (Bouček) stat. rev. * L. sparsibarbis Bouček Lasiochalcida sp. * Neochalcis Kirby Neothybothorax Nikol'skaya N. hetera (Walker) Proconura Dodd P. icongruens (Massi) P. icongruens (Massi) P. ingripes (Fonsolombe) P. persica (Delvare Tamycoryphus Cameron P. infindis Nikol'skaya comb. n. * P. silochalcis Kieffer P. persica (Delvare P. persica (Massi) P. migripes (Fonsolombe) P. persica (Massi) P. silochalcis Kieffer P. subanear (Massi) P. silochalcis (Mass				H. unicolor Walker *	
Dalla Torre K. hofferi (Bouček) Lasiochalcidia Masi L. cincticornis (Walker) L. indescripta (Bouček) stat. rev.* L. sparsibarbis Bouček Lasiochalcidia sp.* Neohoboliorax Nikolškaya N. dertoni (Kieffer) Proconura Dodd Proconura Dodd Prioriginerus (Masi)			Kriechbaumerella	K oracilis (Nikol'skava) comb. n.	
Lasiochalcida Masi Laidescripta (Walker)* Lindescripta (Bouček) stat.rev.* Lasiochalcidia sp.* Neorbybothorax Nikol'skaya N. Jerioni (Kieffer) Proconura Dodd Proconura (Walker) Proconura Dodd Princopynew (Masi) Pincopynew (Poiscolombe) Pincopynew (Poiscolombe) Persica (Delvare Prilochalcis Kieffer Persica (Masi) Persica (M			Della Torra	V. Loffeni (Denžel)	
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P. ligustica (Masi) P. subarea (Masi) P. subarea (Masi) P. subarea (Miss)		Hybothoracini	Psilochalcis Kieffer	P. ceratoniae Delvare	Anomyelois ceratoniae (T.en.: Pyralidae)
				P. ligustica (Masi)	
				P subaenea (Masi)	Carvedon acaciae (Col.: Chrysomelidae)
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Distribution – *Tanycoryphus tibialis* was originally described from Uzbekistan and later reported from Central Asia (Tajikistan & Turkmenistan) (Myartseva & Trjapitzin, 1993). This is first record of the genus from Middle East and Iran. It appears that *T. tibialis* is an endemic species in the Central Asia and some parts of Iran.

Hosts – The specimens were reared from the xylophagous buprestid beetle, *Chrysobothris solieri* Gory & Laporte on pine (*Pinus eldarica*). It is also a known parasitoid of *Chrysobothris affinis* (Fabricius) and *Sphenoptera kaznakovi* Jakovlev (Myartseva & Trjapitzin, 1993).

Remarks – The examination of a type of *V. tibialis*, the type species of the genus *Varzobia* Nikol'skaya, showed than it belongs to the genus *Tanycoryphus* Cameron. *Varzobia* is therefore synonymized here with *Tanycoryphus*, **syn. n.**

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