# Sperm motility parameters of *Barbus barbus callensis* throughout the reproduction season: Computer aided semen analysis and gametes motility duration

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#### **Abstract**

The present study investigated the existing relationship between computer assisted semen motility parameters and gametes motility duration, known to express semen quality and fertilizing capacity in fish. The objective was particularly to identify computer aided semen analysis (CASA) parameters that could be used as potential fertility predictors. Semen samples were collected from the beginning to the end of the spawning season of Barbus barbus callensis, a freshwater fish abundantly distributed throughout North Africa. Semen was simultaneously analyzed using optical microscopy including semen motility duration (SMD) measurement, by the aid of a computerassisted semen analyzer. The measured CASA parameters were: straight line velocity (VSL), average path velocity (VAP), curvilinear velocity (VCL), beat cross frequency (BCF), amplitude of lateral head displacement (ALH), linearity and straightness. The results showed that motility duration evolved in a similar manner as several CASA parameters. Significant correlation coefficients expressed these relationships with r = 0.74, 0.32, 0.16 and 0.45 for VSL, VAP, VCL and BCF, respectively. No correlations were observed when studying relationships between motility duration, STR, LIN and ALH, with r = 0.08, 0.06 and 0.006, respectively. The present results showed that CASA motility parameters are strongly related to motility duration. VSL was revealed as the main parameter being highly correlated to motility duration (r = 0.74). This quantitatively and objectively measured parameter is revealed to be a useful indicator of semen quality and could serve as a potential indicator of fertility outputs in fish.

**Keywords:** Barbus barbus callensis, Reproduction season, Semen motility, CASA.

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#### Introduction

The barbel (*Barbus barbus callensis*) is a freshwater fish inhabiting North Africa (Almaça, 1970). Although, this species has been reported in literature since 1842 (Cuvier and Valenciennes, 1842), little is known concerning the reproduction physiology, particularly concerning variations of semen motility parameters during the reproduction season.

The spermatozoa motility of most teleost fish differs from mammals. Gametes are immotile on ejaculation and the activation is induced only after the delivery of sperm into an aqueous environment or into artificial media (Scott and Baynes, 1980; Perchecpoupard *et al.*, 1997; Kime *et al.*, 2001). In freshwater species, spermatozoa are immotile in relation to the iso-Mackie Building osmolarity of the seminal fluid and a hypo-osmotic shock is required for the initiation of sperm motility (Kime *et al.*, 2001).

quality, key Sperm a factor determining the fertilizing outputs, is significantly related to motility quality (Billard, 1978; Cosson et al., 1991; Lahnsteiner et al., 1997; Fauvel et al., 1999; Au et al., 2002). In fish, it is shown particularly that semen motility duration (SMD) plays a key role in terms of fertility soundness (Billard, 1978: Kozdrowski etal., Gennotte et al., 2012). The more gametes are motile, the more probability to reach the micropyle is important.

However, little is known concerning the variation of sperm motility duration throughout the reproduction season, and in our knowledge no reports have been published concerning existing relationships between SMD and CASA parameters. To date, SMD is still measured using optical microscopy with no automatic alternative. With CASA parameters being measured objectively, existing correlations with SMD could lead to potential fertility predictors in fish.

During the last decades, CASA systems have widely been developed generating series a of including the measurement of the percentage of moving spermatozoa, straight line velocity (VSL, µm/s), average path velocity (VAP, µm/s), curvilinear velocity (VCL, µm/s), beat cross frequency (BCF, Hz), linearity of a curvilinear path (LIN, %), straightness (STR, %), wobble (WOB, %) and amplitude of lateral head displacement (ALH, µm) (Tuset et al., 2008b; Tejerina et al., 2009; Gallego et al., 2013a). These systems are particularly useful in fish where the short duration of motility makes the assessment of sperm quality difficult by direct observation (Kime, 1999; Fauvel et al., 2010).

In different animal species, individual or a group of CASA parameters, including VCL (Larsen *et al.*, 2000), VSL and ALH (Moore and Akhondi, 1996), VSL, VAP and VCL (Viveiros *et al.*, 2010), and BCF (Billard and Cosson, 1992; Oliveira *et al.*, 2013) are suggested as fertility predictors. In fish, only a few studies are dedicated to investigate relationships between CASA

parameters and fertility. However, as in other animal species, conclusions remain controversial with different parameters presented fertility predictors, including VSL, VAP, VCL and BCF (Farrell et al., 1998; Larsen et al., 2000: Fernandez-Santos et al., 2011: Del Olmo et al., 2013; Oliveira et al., 2013). These controversial results are often related to the low number of females finally inseminated.

Based on the presented background, the current study was carried out to explore existing relationships between CASA parameters and motility sperm duration. The experimental protocol consisted of simultaneous semen motility analysis. during the reproduction season of B. barbus callensis, including the measurement of SMD and CASA motility parameters.

### Materials and methods

Fish handling and gamete collection barbel (B.barbus callensis, Cyprinidae family) was collected from Agrioun River (Bejaia), located in Northeastern Algeria (36°36′54.25" N and  $05^{\circ}22'04.33''$  E). Sperm was collected during the spawning season, from June (corresponding to the full reproduction season) to August 2012 (the end of spawning period). The fish were captured using fishing rods connected to a transparent net with 2cm<sup>2</sup> Seventy (70) barbels mesh. assessed during the study period (body weight ranged from 3.85 to 327.70 g and total length from 7.50 to 33.30 cm).

After drying and cleaning the genital papilla with a paper towel to prevent water contamination and initiation of sperm motility, a gentle abdominal pressure was applied to collect semen. The samples contaminated with faeces and urine were discarded (Gallego *et al.*, 2013).

# Semen motility analysis Optical microscopy analysis

Sperm motility was evaluated visually the percentage motile of spermatozoa after activation under  $10 \times 40$ optical microscopy at magnifications. The total duration of semen motility was assessed using a timer immediately after activation; the sperm is considered as immotile when less than 5% sperm remain mobile (Tuset et al., 2008). Semen samples were simultaneously diluted at 1:1000 in fresh river water for motility activation and CASA analysis.

#### CASA motility analysis

The sperm freshwater mixture was analyzed using a Makler cell (Sefi-Medical Instruments, Israel). The CASA system consisted of a triocular optical phase contrast microscope (Nikon, Eclipse E200. Phase contrast 0.90 dry (Japan), equipped with a warming stage at 37°C and a Basler A312fs digital camera (Basler Vision Technologies, Germany). The camera was connected to a computer by an IEEE 1394 interface. Images were captured and analyzed using the Sperm Class Analyzer (SCA, 4.0, 2014) software (Microptic S.L.; Barcelona, Spain). Sampling was conducted using x 10 negative phase contrast objective. Software settings were adjusted to fish sperm. The standard parameter settings were as follows: 25 frames/s; 05–90  $\mu$ m<sup>2</sup> for head area; VCL>10  $\mu$ m/s to classify a spermatozoon as motile.

parameters assessed straight line velocity (VSL), the average velocity measured in a straight line from the beginning to the end of track in um/s, average pathway velocity (VAP), the average velocity of the smoothed cell path in µm/sec, the curvilinear velocity (VCL), the average velocity measured over the actual point-to-point track followed by the cell in µm/sec, the beat cross frequency (BCF), frequency of the sperm head crossing the average path in either direction expressed in Hz, linearity (LIN, %), defined as the ratio VSL/VCL, straightness (STR, %), defined as the ratio VSL/VAP, which expresses the linearity of the average path, and ALH (µm), defined as the amount of lateral displacement of a sperm head along its spatial average trajectory.

# Statistical analysis

Statistical analysis was performed using Stat view 5.0 software (Abaccus). All experiments were repeated at least three times. Values are expressed as mean ± standard error. Coefficients of correlation were used to explore the existing relationships between sperm motility duration (SMD) and CASA parameters. One-way ANOVA followed

by equality of variance F test was used to compare values of each parameter. Differences were considered as statistically significant at p<0.05.

#### Results

# Semen motility duration

Fig. 1 represents the variation of semen motility duration during the study period. A significant increase in semen motility duration was observed from 16  $(64.44 \pm 0.39 \text{ s})$  to 22 June  $(75.12 \pm$ 0.38 s). A regular decrease was subsequently observed from 22 June to 15 July to reach a minimum value of  $21.88 \pm 0.37$  seconds. From 15 July to 6 August, an enhancement of semen motility duration was observed to reach a value of 62 seconds. However, during the two last analyses only three males collected. This were period corresponded to the end of the spawning season.

# CASA motility parameters

Fig. 2 represents the variations in CASA parameters during the study A perfect similarity period. observed when comparing VSL (Fig. 2A) to motility duration (Fig. 1). The maximum and minimum values were reached simultaneously on 22 June and 15 July, with  $18.51\pm0.92$  and  $11.50\pm$ 0.45 µm/s for VSL and 75.12 ± 0.38 and 21.88±0.37 seconds for semen motility duration. This relationship expressed by the highest coefficient of correlation (r = 0.74).

Figs. 2B and 2C represent VAP and VCL variations, respectively.

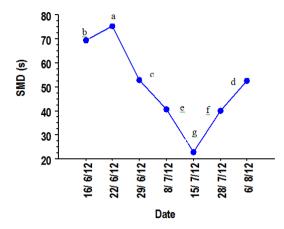


Figure 1: Semen motility duration (SMD) of barbel spermatozoa measured microscopically throughout the reproduction season. Values are expressed as means $\pm$ SE. Different letters (a-g) indicate significant differences between groups on the basis of one-way ANOVA followed by equality of variance F test (p<0.05).

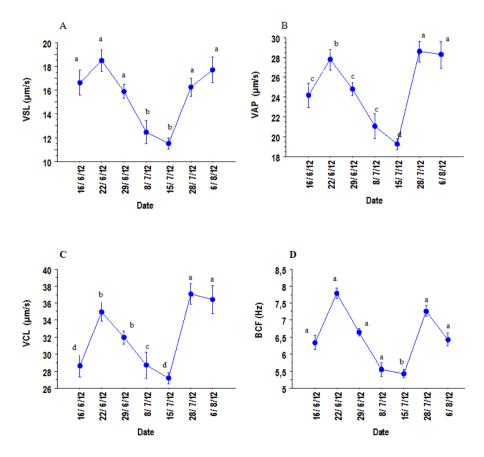


Figure 2: VSL (A), VAP (B), VCL (C) and BCF (D) values during the study period. Values are expressed as means $\pm$ SE. Different letters (a-d) indicate significant differences between groups on the basis of one-way ANOVA followed by equality of variance F test (p<0.05).

VAP is a parameter calculated by smoothing VCL path. The basis of this measurement induced similar evolution (r = 0.95) when comparing these two parameters. The minimum values for VAP and VCL were observed on 15 July at 19.26±0.54µm/s and  $27.15\pm0.68\mu m/s$ , respectively. The maximum values were observed at the end of the reproduction season (28 July) at  $28.61\pm1.03$  µm/s and  $37.11\pm1.23$ μm/s, respectively. Coefficients of correlation with semen motility duration were 0.32 and 0.16, respectively. The beat cross frequency (BCF) (Fig.2D) showed a perfect similarity when compared to VAP and VCL with a simultaneous decrease between 28 July and 6 August. BCF was revealed, to some extent, to be positively correlated to SMD (r = 0.45). ALH, LIN and STR, showed no correlation with SMD (r = 0.006, r = 0.088 and r = 0.068, respectively, data not shown).

#### **Discussion**

Using computer systems, fish semen analysis is achieved readily generating numerous accurate parameters (Wilson-Leedy and Ingermann, 2007; Fauvel et al., 2010). However, it still remains controversial to predict fertility on the basis of CASA analysis (Amann and De Jarnette, 2012; Broekhuijse et al., 2012). fish. semen motility duration, considered as a quantitative objective parameter, is still generated using the conventional microscopic analysis. This parameter is revealed as a potent fertility indicator (Rurangwa et

1998: Kime 2001). al.. etal.. Nevertheless. to knowledge, our relationships with **CASA** motility parameters had never been explored. To investigate these relationships, semen simultaneously analyzed microscopically and automatically by the aid of a CASA system during B. callensis spawning season. The main objective was to explore potential sperm quality and fertility indicators. During the study period, from 16 June to 6 August, semen motility duration showed an average value of  $39.83 \pm 25.24$ seconds (Mean  $\pm$  SD), values similar to those reported for cyprinids (Alavi et al., 2010) and sea bass (Abascal et al., 2007). Seven CASA parameters were measured: VSL, VAP, VCL, ALH, BCF, Linearity and Straightness. The mean recovered values, all samples considered, were 15.13±22.29µm/sec, 24.32±18.82 µm/sec, 31.8±22.29  $\mu$ m/sec, 1.13  $\pm$  0.79 $\mu$ m, 6.46  $\pm$  3.26 Hertz,  $44.86 \pm 27.85 \%$ , and  $56.43 \pm$ 28.05%, respectively. These CASA parameters, presented for the first time, could serve as reference expressing B. barbus callensis sperm motility during the reproduction season. As semen activation was achieved using fresh river water, such references remain of high importance, particularly monitoring the impact of global environment changes on reproduction. CASA motility parameters differ vastly according to the analyzed species. The results showed that values for B. barbus callensis are lower than those reported for commune carp (Cyprinus carpio) but

remain similar to those reported for European eel (Anguilla Anguilla) with  $VSL = 12 \mu m/s$ ,  $VAP = 16 \mu m/s$  and VCL =  $40 \mu m/s$  (Asturiano *et al.*, 2004). Computer aided semen analysis has been used extensively in different domains in evaluating sperm quality in fish (Christ et al., 1996; Alavi and Cosson, 2005). However, it is still difficult to predict fertility outcome in accurate manner (Amann and Dejarnette, 2012; Broekhuijse et al., 2012), mostly due to the deficiency of validated studies confronting CASA parameters and fertility outcomes. This remains difficult even in species such as human and bovine, where CASA is routinely used with continuous fertility feed-back after artificial insemination (Andersson et al., 1992; Barrat et al., 1993; Irvine et al., 1994; Januskauskas et al., 2000). Nevertheless, individual or a group of CASA parameters are reported as correlating significantly with fertility. Thus, it has been shown in rats that fertilizing capacity of spermatozoa correlates with straight line velocity (VSL) (Moore and Akhondi, 1996). In bulls (Farrell et al., 1998; Kathiravan et al., 2011; Oliveira et al., 2013) and humans (Barrat et al., 1993; Irvine et al., 1994; Krause, 1995; Larsen et al., 2000; Hirano *et al.*, 2001), various parameters including VSL, VAP, VCL, BCF and ALH are reported as fertility predictors. In fish, only a few studies have been dedicated to investigate relationships between CASA parameters and fertility, in which contradictory results have been obtained. Thus, VSL,

VAP, VCL and BCF are separately suggested to be correlated with fertility (Billard and Cosson, 1992; Lahnsteiner, 2000; Rurangwa et al., 2004; Viveiros et al., 2010). In fish, the identification of computer parameters that are strongly correlated with sperm quality and fertility is still pending. Alternative to fertility exploration, the present study was conducted to correlate CASA with parameters semen motility duration. When studying the evolution of semen motility duration throughout the reproduction season, no correlation had been observed with linearity, straightness and ALH (r = 0.06, 0.07and 0.005, respectively). On the other hand, a perfect similarity was observed with VSL, VAP, VCL and BCF expressed by high coefficient of correlations (r = 0.74, 0.33, 0.16 and 0.44, respectively). The progressive movement, measured microscopically, is one of the most obvious parameters used in the assessment of sperm quality in fish (Kime et al., 2001). Its closer equivalent in CASA parameters is the straight line velocity (VSL). VSL corresponds to the average velocity measured by considering a straight line distance from the beginning to the end of spermatozoa track. It indicates the aptitude of the gametes to move forward in a straight line. This indicates that semen motility duration, expressing the aptitude of the gametes to be motile for a long period, expresses also the quality of the gametes motility to progress in a straight line, increasing thereby the probability to meet the oocytes.

Generally, the present study pointed out the interest of **CASA** motility parameters as potent indicators of sperm quality in fish. Significant correlations were established with semen motility duration, particularly concerning VSL. As VSL is reported previously in different animal species as a fertility indicator, this parameter could offer a real opportunity in fish. However, the present results need to be strengthened studies including by further simultaneous analysis of semen motility duration, CASA parameters and fertility outcomes.

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# References

- **Abascal, F.J., Cosson, J. and Fauvel, C., 2007.** Characterization of sperm motility in sea bass: the effect of heavy metals and physicochemical variables on sperm motility. *Journal of Fish Biology*, 70, 509–522.
- Alavi, S.M.H. and Cosson, J., 2005. Sperm motility in fishes: (I) effect of temperature and pH. *Cell Biology International*, 29, 101–10.
- Alavi, S.M.H., Rodina, M., Hatef, A., Stejskal, V., Policar, T., Hamačkova, J. and Linhart, O., 2010. Sperm motility and monthly variations of semen characteristics in *Perca fluviatilis* (Teleostei:

- Percidae). Czech Journal of Animal Science, 55, 174–182.
- Almaça, C., 1970. Sur les barbeaux (Genre et sous-genre Barbus) de l'Afrique du Nord. *Bulletin du Muséum National d'histoire Naturelle*, 42(1), 141–158.
- Amann, R.P. and Dejarnette, J.M., 2012. Impact of genomic selection of AI dairy sires on their likely utilization and methods to estimate fertility: aparadigm shift. *Theriogenology*, 77, 795–817.
- Andersson, M., Hellman, T., Holmstrom, B.G. and Jokinen, L., 1992. Computerized and subjective assessments of post-thaw motility of semen from Finnish Ayrshire AI bulls in relation to non-return rates. *Acta Veterinaria Scandinavica*, 33, 89–93.
- Asturiano, J.F., Perez, L., Garzon, D.L., Marco-jimenez, F., Penaranda, D.S. and Vicente, J.S., 2004. Physio-chemical characteristics of seminal plasma and development of media and methods for the cryopreservation of European eel sperm. *Fish Physiology and Biochememistry*, 30, 283–293.
- Au, D.W.T., Chiang, M.W.L., Tang, J.Y.M., Yuen, B.B.H., Wang, Y.L. and Wu, R.S.S., 2002. Impairment of sea urchin sperm quality by UV-B radiation: predicting fertilization success from sperm motility. *Marine Pollution Bulletin*, 44, 583–589.
- Barrat, C.L.R., Tomlinson, M.J. and Cook, I.D., 1993. Prognostic significance of computerized

- motility analysis for in vivo fertility. *Fertility and Sterility*, 60, 520–525.
- **Billard, R., 1978.** Changes in structure and fertilizing ability of marine and freshwater fish spermatozoa diluted in media of various salinities. *Aquaculture*, 14, 187–198.
- **Billard, R. and Cosson, J., 1992.** Some problems related to the assessment of sperm motility in freshwater fish. *Journal of Experimental Zoology*, 261, 122–131.
- Broekhuijse, M.L., Sostaric, E., Feitsma, H. and Gadella, B.M., 2012. Application of computer-assisted semen analysis to explain variations in pig fertility. *Journal of Animal Science*, 90, 779–89.
- Christ, S.A., Toth, G.P., Mccarthy, H.W., Torsella, J.A. and Smith, M.K., 1996. Monthly variation in sperm motility in common carp assessed using computer-assisted sperm analysis (CASA). *Journal of Fish Biology*, 48, 1210–1222.
- Cosson, J., Billard, R., Redondomuller, C. and Cosson, M.P., 1991.

  In vitro incubation and maturation of carp (Cyprinus carpio) spermatozoa.

  Bulletin of the Institute of Zoology.Academia Sinica Monograph, 16, 249–261.
- **Cuvier, G. and Valenciennes, A., 1842.** Histoire naturelle des Poissons. Paris: *P. Bertrand.* pp. 1828–1850.
- Del olmo, E., Bisbal, A., Marotomorales, A., García-alvarez, O., Ramon, M., Jimenez-rabadan, P., Martínez-pastor, F., Soler, A.J., Garde, J.J. and Fernandez-santos,

- **M.R., 2013.** Fertility of cryopreserved ovine semen is determined by sperm velocity. *Animal Reproduction Science*, 138, 102–109.
- Farrell, P.B., Presicce, G.A., Brockett, C.C. and Foote. **R.H.**. 1998. Ouantification of bull sperm characteristics measured bv computer-assisted sperm analysis (CASA) and the relationship to fertility. Theriogenology, 48, 871-879.
- Fauvel, C., Savoye, O., Dreanno, C., Cosson, J. and Suquet, M., 1999. Characteristics of sperm of captive seabass in relation to its fertilization potential. *Journal of Fish Biology*, 54, 356–369.
- **Fauvel, C., Suquet, M. and Cosson, J., 2010.** Evaluation of fish sperm quality. *Journal of Applied Ichthyology*, 26, 636–643.
- Fernandez-santos, M.R., Soler, A.J., Ramón, M., Ros-santaella, J.L., Maroto-morale, A., García-alvarez, O., Bisbal, A., Garde, J.J. and Santiago-moreno, J., 2011. Effect of post-mortem time on post-thaw characteristics of Spanish ibex (Capra pyrenaica) spermatozoa. Animal Reproduction Science, 129, 56–66.
- Gallego, V., Pérez, L., Asturiano, J.F. and Yoshida, M., 2013. Relationship between spermatozoa motility parameters, sperm/egg ratio, and fertilization and hatching rates in pufferfish (*Takifugu niphobles*). *Aquaculture*, 416–417, 238–243.

- Gallego, V., Pérez, L., Asturiano, J.F. and Yoshida, M., 2013a. Study of pufferfish (*Takifugu niphobles*) sperm: development of methods for short-term storage, effect of different activation media and role of intracellular changes in Ca<sup>2+</sup> and K<sup>+</sup> in the initiation of sperm motility. *Aquaculture*, 414–415, 82–91.
- Gennotte, V., Francois, E., Rougeot, C., Ponthier, J., Deleuze, S. and Mélard, C., 2012. Sperm quality analysis in XX, XY and YY males of the Nile tilapia (Oreochromis niloticus). Theriogenology, 78, 210–217.
- Hirano, Y., Shibahara, H., Obara, H., Suzuki, T., Takamizawa, S., Yamaguchi, C., Tsunoda, H. and Sato, I., 2001. Relationships between sperm motility characteristics assessed by the Computer-Aided Sperm Analysis (CASA) and fertilization rates In vitro. *Andrology*, 18, 213–218.
- Irvine, D.S., Macleod, I.C., Templeton, A.A., Masterton, A. and Taylor, A., 1994. A prospective clinical study of the relationship between the computer-assisted assessment of human semen quality and the achievement of pregnancy in vivo. *Human Reproduction*, 9, 2324–2334.
- Januskauskas, A., Johannisson, A., Soderquist, L. and Rodriguezmartinez, H., 2000. Assessment of sperm characteristics post-thaw and response to calcium ionophore in relation to fertility in Swedish dairy

- AI bulls. *Theriogenology*, 53, 859–875.
- Kathiravan, P., Kalatharan, J., Karthikeya, G., Rengarajan, K. and Kadirvel, G., 2011.

  Objective sperm motion analysis to assess dairy bull fertility using computer- aided system.

  Reproduction in Domestic Animals, 46, 165–172.
- **Kime, D.E., 1999.** A strategy for assessing the effects of xenobiotics on fish reproduction. *The Science of the Total Environment*, 225, 3–11.
- Kime, D.E., Van look, K.J.W., Mcallister, B.G., Huyskens, G., Rurangwa, E. and Ollevier, F., 2001. Computer assisted sperm analysis (CASA) as a tool for monitoring sperm quality in fish. Comparative biochemistry and physiology, 130, 425–433.
- **Kozdrowski, R., Dubiel, A., Bielas, W.** and Dzieciol, M., 2007. Two protocols of cryopreservation of goat semen with the use of computerassisted semen analysis system. *Acta Veterinaria Brno*, 76, 601–604.
- **Krause, W. 1995.** Computer-assisted semen analysis systems: comparison with routine evaluation and prognostic value in male fertility and assisted reproduction. *Human Reproduction*, 10, 60–66.
- Lahnsteiner, F., Berger, B., Weismann, T. and Patzner, R.A., 1997. Sperm motility and seminal composition in the turbot (*Lota lota*). *Journal of Applied Ichthyology*, 13, 113–119.

- **Lahnsteiner, F. 2000.** Semen cryopreservation in the Salmonidae and in the Northern pike. *Aquaculture Research*, 31, 245–258.
- Larsen, L., Scheike, T., Jensen, T.K., Bonde, J.P., Ernst, E., Hjollund, N.H., Zhou, Y., Skakkebaek, N.E. and Giwercman, A., 2000. Computer-assisted semen analysis parameters are predictors for fertility of men from the general population *Human Reproduction*, 15, 1562–1567.
- Moore, H.D.M. and Akhondi, M.A., 1996. Fertilizing capacity of rat spermatozoa is correlated with decline in straight line velocity measured by continuous computeraided sperm analysis. *J. Andrology*, 17, 50–60.
- Oliveira, L.Z., Arruda, R., Andrade, A., Celeghini, E., Reeb, P., Martins, J., Santos, R., Beletti, M., Perew, R., Monteiro, F. and Lima, V., 2013. Assessment of in vitro characteristics and their importance in the prediction of conception rate in a bovine timed-AI program. *Animal Reproduction Science*, 137, 145–155.
- Perchec-poupard, G., Gatti, G.L., Cosson, J., Jeulin, C., Fierville, F. and Billard, R., 1997. Effects of extracellular environment on the osmotic signal transduction involved in activation of motility of carp spermatozoa. *Journal of Reproduction and Fertility*, 110, 315–327.
- Rurangwa, E., Roelants, I., Huyskens, G., Ebrahimi, M., Kime, D.E. and

- Ollevier, F., 1998. The minimum effective spermatozoa: egg ratio for artificial insemination and the effects of mercury on sperm motility and fertilisation ability in *Clarias gariepinus*. *Journal of Fish Biol*ogy, 53, 402–413.
- Rurangwa, E., Kime, D.E., Ollevier, F. and Nash, J.P., 2004. The measurement of sperm motility and factors affecting sperm quality in cultured fish. *Aquaculture*, 234, 1–28.
- **Scott, A.P. and Baynes, S.M., 1980.** A review of the biology, handling and storage of salmonid spermatozoa. *Journal of Fish Biology*,, 17, 707–739.
- **Tejerina, F., Morrell, J., Petterson, J., Dalin, A.M. and Rodriguez- martinez, H., 2009.** Routine assessment of motility of ejaculated stallion spermatozoa using a novel computer-assisted motility analyser (Qualisperm<sup>TM</sup>). *Animal Reproduction Science.*, 6, 380–385.
- Tuset, V.M., Dietrich, J., Wojtczak, M., Słowinska, M., demonserrat, J. and Ciereszko, A., 2008. Relationships between motility morphology, and fertilization capacity in rainbow trout (Oncorhynchus mykiss) spermatozoa. Journal of Applied Ichthyology, 24, 393-397.
- Tuset, V.M., Trippel, E.A. and demonserrat, J., 2008b. Sperm morphology and its influence on swimming speed in Atlantic cod.

*Journal of Applied Ichthyology*, 24, 389–405.

Viveiros, A.T.M., Nascimento, A.F., Orfão, L.H. and Isaú, Z.A., 2010. Motility and fertility of the subtropical freshwater fish streaked prochilod (*Prochilodus lineatus*) sperm cryopreserved in powdered

coconut water. *Theriogenology*, 74, 551–556.

**Wilson-leedy, J.G. and Ingermann, R.L., 2007.** Development of novel CASA system based on open source software for characterization of zebrafish sperm motility parameters. *Theriogenology*, 67, 661–672.