

Karyotype Analysis of Populations in Five *Satureja* Species from Iran

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ABSTRACT

A karyological study of five taxa (15 populations) of the genus *Satureja* L. from different geographic origins is presented. The haploid levels were different among species ($n=12$ and 15). The work deals with chromosome number and morphometry. We found two of the usual basic chromosome numbers in this study for *satureja* species, $x=12$ and $x=15$. All populations were diploid and were located in 1B or 2B Stebbins classes. The karyotypic results showed the diversity among the species as mostly displayed median region (m) and submedian region (sm) and the chromosome lengths were determined between $0.60\ \mu\text{m}$ in *Satureja sahendica* Bornm. (1041) to $2.27\ \mu\text{m}$ in *S. macrantha* C.A.Mey. (3). Detailed karyotype analysis allows us to group the different populations and determine the relationships among them. It also pointed out the possibility of inter and intra-species cross to improve plants.

INTRODUCTION

The genus *Satureja* is one of the most important genera of the Lamiaceae family. It is described in the Menthaeae tribe (sub-tribe Menthinae) belonging to the Nepetoideae subfamily and includes about 480 species in the world [1]. This genus is represented in Iranian flora by 16 species, 10 of which are endemic for the country (*S. sahendica* Bornm., *S. edmondii* Briquet, *S. kallarica* Jamzad, *S. kermanshahensis* Jamzad, *S. khuzistanica* Jamzad, *S. bachtiarica* Bunge, *S. intermedia* C. A. Mey., *S. isophylla* Rech.f., *S. atropatana* Bunge and *S. rechingeri* Jamzad [2].

Satureja species are native to warm temperate regions and may be annual or perennial. They are subshrubs and low-growing herbs, reaching heights of 20- 50 cm, with flowers forming in whorls on the stem, white to pale pink-violet. They are commonly

well known by native residents and are used by native inhabitants as medicinal plant, spice or source of essential oils [3-5].

In this study five different (four of which were endemic) taxa (15 populations) of the *Satureja* species including *S. macrantha* C. A. Mey, *S. sahendica*, *S. edmondii*, *S. kermanshahensis* and *S. khuzistanica* were examined.

S. macrantha is a small shrub distributed in western and northwestern parts of Iran. *S. sahendica* is a late flowering species (late summer and fall), growing on rock walls and rocky slopes and distributed in western and northwestern parts of Iran [6]. *S. edmondii* is mainly perennials and occupy mountainous habitats in northern and western parts of Iran. *S. khuzistanica*, is one of the important medicinal plants among the nomadic residents of southwestern of Iran and grows on dry, limestone

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rocky slopes poor in mineral content. This is a traditional herbal that uses a decoction of the aerial parts (mainly dried leaves) as toothpaste and oral disinfectant [7]. Finally, *S. kermanshahensis* species grows in crevices of rocks in the Kermanshah province in the west of Iran.

A large number of samples in each species and their drug trustees increase the importance of diversity studies in this genus and species. Within the *Satureja* genus the genetic diversity has been investigated using morphological characters [8,9] Enzyme electrophoresis [10,11] and molecular markers such as RAPD, SAMPL and AFLP [11-16]. Studying karyotypes and knowing the number of chromosomes is not only useful in predicting morphological similarity and diversity among *Satureja* species, but they are also valuable sources of taxonomic and biosystematics information.

The basic chromosome numbers that have been reported for the *Satureja* species is vary as follows: *S. multiflora*, x= 6 [17]; *S. acinos*, x=9 [18]; *S. vulgaris*, x= 10 [18]; *S. bulgarica* and *S. spicigera*, x=11, [19,20]; *S. macrosiphonia*, x=12 [20]; *S. mutica*, x=13 [20]; *S. sahendica*, x=14 [20]; *S. cristata*, *S. cuneifolia*, *S. bakhtiarica*, *S. rechingeri*,

S. spicigera, *S. montana*, x= 15 [19, 21-23]; *S. robusta*, x= 21 [24] and *S. hortensis*, x= 24 [25].

There was rarely a report to date on the detailed karyotype of these species in this research, also different and even inaccurate reports on the number of chromosomes of some of the studied species have been observed. Therefore, we have been tried to resolve the ambiguities in this research. Thus the karyotype analysis of some species in this report was the first time counted in Iran. The objectives of this study are 1) to present karyological information, particularly the differences among them and 2) to determine the chromosome number and haploidy levels of these taxa.

MATERIAL AND METHODS

Plant materials

The materials used in this study were collected in different areas of Iran. The localities, herbarium codes, geographical situation and species names are shown in Table 1. Vouchers are deposited in gene bank RIFR (Research Institute of Forest and Rangelands from Iran).

Table 1 Geographical characteristics of *Satureja* species localities

species	localities	Herbarium Code (RIFR)	Geographical Status	Altitude (m, a.s.l)
<i>S. macrantha</i> C.A.Mey	Arasbaran	3	Northwest of Iran, North of East Azerbaijan province	536
<i>S. macrantha</i> C.A.Mey	Azerbayejan1	4	Northwest of Iran	657
<i>S. macrantha</i> C.A.Mey	Azerbayejan2	17	Northwest of Iran	832
<i>S. sahendica</i> Bornm.	Bijar	1041	Northeast of Kordestan province	2200
<i>S. sahendica</i> Bornm.	Gheydar	1042	South of Zanjan province	2470
<i>S. sahendica</i> Bornm.	Golabir	1043	Southwest of Zanjan province	1805
<i>S. sahendica</i> Bornm.	Soltanieh1	1044	South of Zanjan	1919
<i>S. sahendica</i> Bornm.	Soltanieh2	1045	South of Zanjan	2032
<i>S. sahendica</i> Bornm.	Sahand	7	South of Tabriz city	2200
<i>S. sahendica</i> Bornm.	Songhor	8	Northeast of Kermanshah	2470
<i>S. edmondi</i> Briq.	Kamyara	770	South of Sanandaj	1320
<i>S. edmondi</i> Briq.	Perav mountain	771	North of Kermanshah	1480
<i>S. edmondi</i> Briq.	Songhor	772	Northeast of Kermanshah	1365
<i>S. kermanshahensis</i> Jam.	Sumar	601	West of Kermanshah province	1180
<i>S. khuzistanica</i> Jam.	Lorestan	705	West of Iran	486

Root tip meristems obtained from seedlings were pretreated with 0.5% saturated α -Bromo naphthalene at 40°C for 2.5h, fixed in 40% formaldehyde and 1% chromic acid (1:1) for at least 15 h at room temperature, then root tips were rinsed for 1 h in tap water. Hydrolysis was carried out with 1N NaOH at 60 °C for nine minutes, dyed with Aceto- Iron-hematoxylin for 7-8h and squashed in a droplet of 45% acetic acid and lactic acid (10:1). Each of the prepared samples was observed with an optical microscope (BH2Olympus supplemented digital color video camera) at a magnification of 2020x. The best metaphase plates were selected and measured by Micro measure 3.3 software [26]. In each mitotic metaphase (at least five or six plates) the arm's length of each chromosome was measured and all parameters were estimated to characterize the karyotypes, according to the previous studies [20, 27-32].

Karyotype asymmetry was estimated by four different methods namely, the difference of relative length (DRL); total form percentage (TF%) [33]; intra-chromosomal asymmetry index (A_1) and inter-chromosomal asymmetry index (A_2) [34]. Both indices (A_1 and A_2) are independent of chromosome number and size. Dispersion index (DI) [35] and also karyotypic evolution by using the symmetry classes of Stebbins (SC) [36] has been determined. The karyotype formula was determined according to the classification of Levan [37]. For each population, Metaphase plates and karyogram of somatic chromosomes were adjusted based on length of chromosome size (arranged largely to small).

To determine the diversity between populations, one-way balanced ANOVA was performed on normal data and parameters mean were compared by Duncan's test. To evaluate the contribution of each karyotypic parameter to the ordination of species, principal component analysis (PCA) was performed.

Clustering was performed using Ward's cluster analysis method after calculation of the Cophenetic correlation coefficient (r) to examine karyotype similarity among populations. Numerical analyses were performed using JMP ver. 3.1.2, 1995 [38]; StatistiXL ver 1.8, 2007 [39] and SAS ver. 6.12, 1996 [40] soft wares.

RESULTS

With relation to the plant materials assayed, Karyotype analyses of five species (15 populations) of *Satureja* were determined. Somatic cells of investigated species had $2n=2x=24$ chromosomes for *S. kermanshahensis*; *S. macrantha* and $2n=2x=30$ chromosomes for *S. sahendica*; *S. edmondi* and *S. khuzistanica* species. Metaphase plates and karyogram of somatic chromosomes for five species are illustrated in Figure 1. The average total chromosomal length of investigated species ranged from 0.60 to 2.27 μm . The somatic chromosome numbers ($2n$), ploidy levels, ranges of

chromosome length, symmetry index percentage, intra and inter-asymmetry indices, the difference of range relative length, total form percentage, symmetry classes, total karyotype length and karyotype formula of the investigated taxa and populations are summarized in (Table 2).

The mean value of the chromosome's long arm was varied from 0.52 μm in *S. edmondi* (770) to 0.76 μm in *S. edmondi* (771). The averages of the chromosome's short arm were different from 0.39 μm in *S. sahendica* (1041) to 0.63 μm in *S. edmondi* (771). The mean value of chromosome's total length was varied from 0.95 μm in *S. edmondi* (770) to 1.39 μm in *S. edmondi* (771) and finally, the mean value of chromosome's arm ratio was changing from 1.21 in *S. edmondi* (770) to 1.69 in *S. sahendica* (7) (Table 4).

All chromosomes were metacentric (m) in populations of *S. macrantha* (17) and *S. edmondi* (770,771 and 772) while some chromosomes in populations of *S. macrantha* (3 and 4), *S. sahendica* (1041, 1042, 1043, 1044, 1045, 7 and 8), *S. kermanshahensis* and *S. khuzistanica* were metacentric (m) or sub-metacentric (sm) (Table 2). The dispersion index (DI) is calculated as a proportional measurement of the centromeric gradient to the coefficient of variation for chromosome length. The highest value of DI was found in *S. macrantha* (17) (14.81) and the lowest value of DI was found in *S. sahendica* (1042) (7.62) (Table 4). Symmetry type of (Stebbins 1971) [36] and asymmetry indices of (Romero-Zarco 1986) [34] are given in (Table 2). In terms of the Stebbins' system, the karyotype of populations seizes 1B and 2B classes, which are considered majorly primitive classes in this system. All populations of *S. macrantha*, *S. sahendica*, *S. edmondi* and *S. kermanshahensis* are classified as 1B group except of *S. macrantha* (3), *S. sahendica* (7) and *S. khuzistanica* species that stand as 2B category (Table 2).

Using the Romero-Zarco asymmetry indices of A_1 and A_2 we determined the more asymmetric karyotype among the populations which have similar Stebbins classes of symmetry. In the populations with 2B classes, *S. sahendica* (7) possesses the highest A_1 value (0.38) and the lowest TF% value (37.47). In the populations with 1B class, *S. edmondi* (770) possesses the lowest A_1 value (0.16) and the highest TF% value (45.40) (Table 2).

To analyze the variability of the karyotypes among populations, length of the chromosome, long and short arms of the chromosome, arm ratio values, centromeric index, long and short arm percent, relative length percent, the difference of relative length, total form percentage and intra-chromosome asymmetry index were compared by one-way analysis of variance based on completely randomized design (CRD). Also, the Duncan test was carried out to test the differences between each

pair of means. The results of variance analysis revealed significant differences between the populations based on eleven karyotypic characteristics ($p < 0.05$ and $p < 0.01$) (Table 3). The Duncan test applied to the chromosome morphometric traits showed a highly significant difference among all examined populations (Table 4). Therefore, mean values of chromosome total length varied from 1.40 μm in populations *S. edmondi* (771) to 0.95 μm in populations *S. edmondi* (770).

The mean values of chromosome long arms varied from 0.764 μm in *S. edmondi* (771) to 0.518 μm in *S. edmondi* (770). Also, the mean values of chromosome short arms were different from 0.632 μm in *S. edmondi* (771) to 0.365 μm in *S. sahendica* (7). Using principal components analysis (PCA), the first two independent components accounted for about 81.28% of the total variation.

The first component emphasized total length, long and short arms, arm ratio, centromere index, short arm percent, total form percentage and intra-chromosome asymmetry index which had the highest coefficients of Eigenvectors and were important characters for classification of populations with about 50.24% of total variation. Long arm percent, relative length percent and difference of relative length were important traits in the second component (31.04%) (Table 5).

Grouping of investigated populations was based on their karyotypic traits (Fig. 3). The results showed that populations of *Satureja* genus have been grouped in a separate cluster.

By cutting the dendrogram resulting from cluster analysis by Ward's method with cophenetic correlation coefficient ($r = 0.85$) with a metric distance of 3.11, the populations were classified into six groups.

The highest metric distance (8.99) was obtained between *S. macrantha* (3) and *S. sahendica* (1041), which implies the least affinity between them. The lowest metric distance (0.55) was obtained between *S. sahendica* (1041) and *S. khuzistanica*, which implies the least karyotypic difference between them. Due to the single population of the two species *S. kermanshahensis* and *S. khuzistanica*. In this paper, their position in the cluster (Table 6) is based on the similarity of chromosome number and

chromosome size are located next to the *S. macrantha* and *S. sahendica* species (Fig. 3).

The diagram of population dispersion, based on the first two components, showed that the populations were separated into six groups, which completely fits with results obtained through the grouping analysis by Ward's method (Fig. 2). In Figure 2, as can be seen, the species are classified into different categories and separated from each other based on about 81% of variance and the coefficients of the first and second components. *S. edmondi* are located in the lowest right area of the plot due to factors such as TL, LA, SA, AR, CI, SA%, TF% (based on traits with high coefficients in the first component compared to the second component). Also, *S. macrantha*, due to the relatively high trait coefficients in both, the first and second components, are located on the right and at the top of the plot. *S. sahendica* and *S. khuzistanica* are located on the left side of the plot according to the traits of LA%, RL % and DRL (traits with higher coefficients in the second component than the first component) (Table 5, Fig. 2). Populations of *S. macrantha* (17) and *S. edmondi* (770) are classified as separate groups. Also three populations of *S. macrantha* (3), *S. macrantha* (4) and *S. kermanshahensis* together they formed a separate group, three populations of *S. sahendica* (1041), *S. sahendica* (7) and *S. khuzistanica* together they formed other separate group.

Two populations of *S. edmondi* (771) and *S. edmondi* (772) together they formed a separate group and finally, the remaining five *S. sahendica* (8, 1042, 1043, 1044 and 1045) together they were classified into separate groups. The population of *S. macrantha* (17) due to having the highest values of SA%, RL%, TF% and DI and the population of *S. edmondi*(770) due to having the highest values of CI, TF% and the lowest values of TL, LA, AR and A_1 were grouped into separate classes. The populations of *S. sahendica* (7 and 1041) were isolated from other populations of *S. sahendica* due to having significant differences in the amount of AR, TF% and DRL values and were grouped with *S. khuzistanica* species in the same cluster. The population of *S. edmondi* (770) has been separated from other populations of *S. edmondi* due to having significant differences in TL, LA and SA values (Table 4; Fig. 2).

Table 2 Somatic chromosome number (2n), ploidy level, ranges of chromosome length, Arm ratio (AR), asymmetry indices of Romero Zarco (A₁, A₂), Dispersion Index(DI), difference of range relative length (DRL), total form percentage (TF %), symmetry classes (SC) of Stebbins, total karyotype length (TKL) and Haploid karyotype formula (K.F.) (m: metacentric, sm: submetacentric)

Population	2n	Ploidy level	Chromosome length range	AR	A ₁	A ₂	DI	DRL	TF%	SC	TKL (μm)	K.F.
<i>S. macrantha</i> (3)	24	2x	0.72-2.27	1.51	0.35	0.37	14.49	10.45	38.90	2B	28.92	10m + 2sm
<i>S. macrantha</i> (4)	24	2x	0.83-1.99	1.37	0.25	0.30	12.46	8.35	42.54	1B	27.80	11m + 1sm
<i>S. macrantha</i> (17)	24	2x	0.73-2.21	1.26	0.19	0.34	14.81	9.60	44.75	1B	30.86	12m
<i>S. sahendica</i> (1041)	30	2x	0.60-1.57	1.60	0.33	0.28	9.98	6.38	40.32	1B	30.52	10m + 5sm
<i>S. sahendica</i> (1042)	30	2x	0.83-1.66	1.52	0.33	0.19	7.62	4.95	40.05	1B	34.50	12m + 3sm
<i>S. sahendica</i> (1043)	30	2x	0.78-1.67	1.45	0.29	0.22	8.94	5.40	40.95	1B	34.44	14m + 1sm
<i>S. sahendica</i> (1044)	30	2x	0.76-1.70	1.47	0.31	0.26	9.52	5.84	40.90	1B	32.10	12m + 3sm
<i>S. sahendica</i> (1045)	30	2x	0.81-1.75	1.58	0.33	0.23	8.80	5.45	40.01	1B	34.06	13m + 2sm
<i>S. sahendica</i> (7)	30	2x	0.66-1.58	1.67	0.38	0.27	9.97	6.28	37.47	2B	29.10	8m + 7sm
<i>S. sahendica</i> (8)	30	2x	0.77-1.67	1.51	0.32	0.24	8.90	5.42	40.56	1B	32.88	12m + 3sm
<i>S. edmondi</i> (770)	30	2x	0.64-1.59	1.21	0.16	0.27	11.31	6.68	45.40	1B	28.48	15m
<i>S. edmondi</i> (771)	30	2x	0.92-2.07	1.22	0.17	0.23	10.04	5.58	45.20	1B	41.90	15m
<i>S. edmondi</i> (772)	30	2x	0.89-1.99	1.26	0.20	0.21	8.67	5.73	44.66	1B	38.34	15m
<i>S. kermanshahensis</i> (601)	24	2x	0.68-1.68	1.41	0.29	.31	13.17	7.83	41.89	1B	25.54	11m + 1sm
<i>S. khuzistanica</i> (705)	30	2x	0.71-1.75	1.57	0.34	0.27	9.21	6.68	39.43	2B	31.62	10m + 5sm

Table 3 The results of analysis of variance for karyotypic data based on CRD design

S.O.V	D.F.	TL	LA	SA	AR	CI	LA%	SA%	RL%	DRL	TF%	A ₁	A ₂	DI
Population	14	0.45 **	0.011 **	0.016 **	0.069 **	0.002 **	0.430 **	0.384 **	1.348 **	8.058 *	18.628 **	0.014 **	0.006 ^{ns}	0.323 ^{ns}
Error	30	0.011	0.003	0.002	0.015	0.0003	0.017	0.019	0.0147	3.760	3.536	0.002	0.004	0.169
%C.V.		9.23	8.99	11.04	8.56	4.58	3.24	4.95	1.76	13.90	4.53	8.92	13.02	12.83

** and * significant at 1% and 5% levels of probability respectively; ^{ns}: no significant

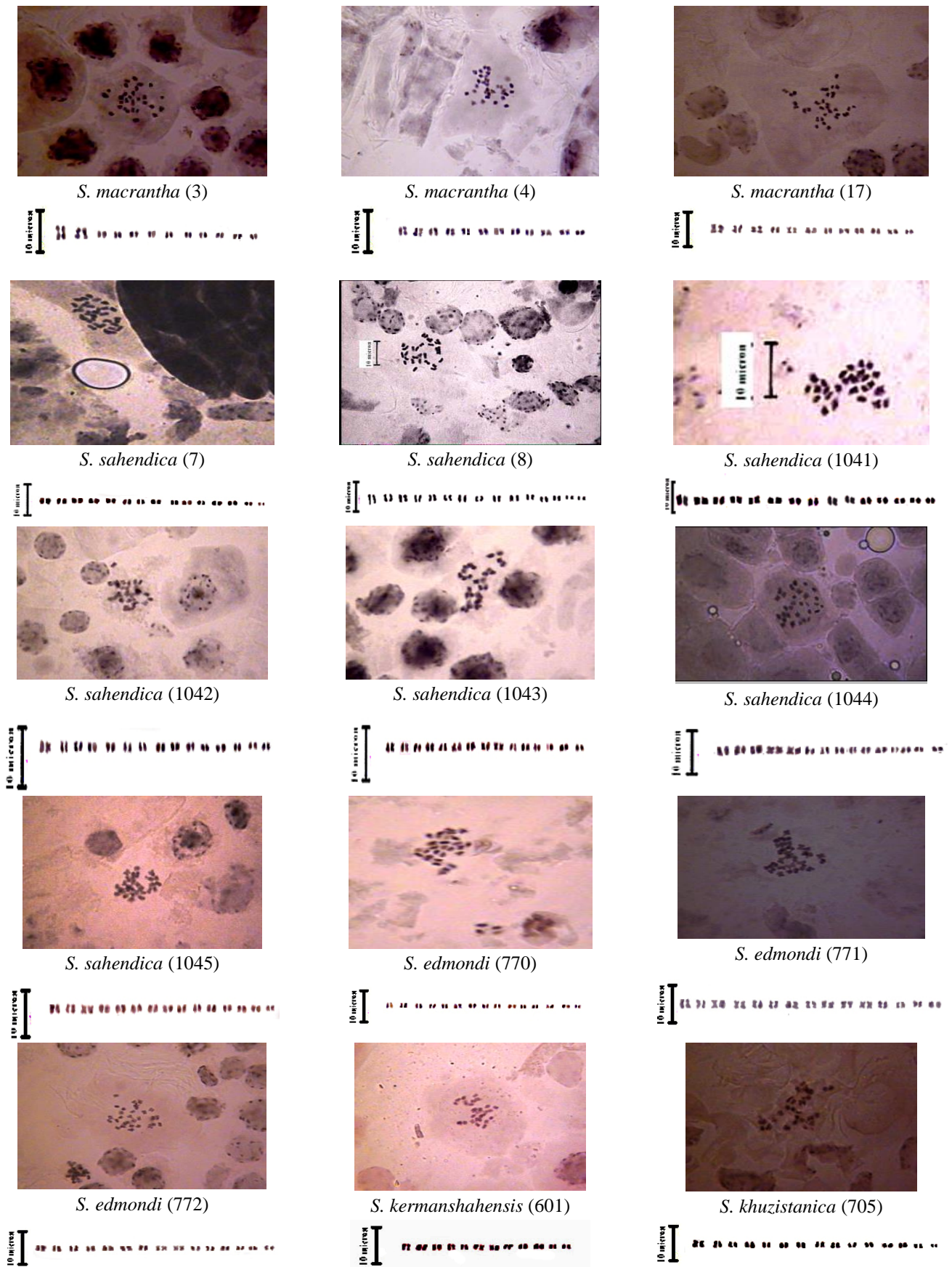


Fig. 1 Mitotic metaphase of *Satureja* populations accompanied by karyograms.

Table 4 Mean of parameters of chromosomes analysis of *Satureja* populations. TL: total length of chromosome, LA: long arm, SA: short arm, AR: arm ratio, CI: Centromeric index, LA%: long arm percent, SA%: short arm percent, RL%: Relative length percent; TF%: total form percentage, DRL: difference of relative length, A₁: intrachromosome asymmetry index, A₂: interchromosome asymmetry index, DI: Dispersion Index

In each column the same letters are not significantly different at $P \leq 0.05$

Population	TL	LA	SA	AR	CI	LA%	SA%	RL%	TF%	DRL	A1	A2	DI
<i>S. macrantha</i> (3)	1.205 bc	0.719ab	0.486bc	1.507 ab	0.396cde	4.690a	3.116bc	7.862a	38.909bc	10.446a	0.349ab	0.369a	14.492 ab
<i>S. macrantha</i> (4)	1.158 bcd	0.665ab	0.492bc	1.370bcde	0.419abc	4.615ab	3.368a	8.028a	42.537ab	8.346abc	0.253bc	0.304abc	12.464abc
<i>S. macrantha</i> (17)	1.286 ab	0.710ab	0.575ab	1.260cde	0.440ab	4.402b	3.496a	7.928a	44.753a	9.604 ab	0.193cd	0.336ab	14.807 a
<i>S. sahendica</i> (1041)	1.017 cde	0.621bc	0.396cd	1.605ab	0.381de	3.925c	2.450d	6.418b	40.320bc	6.378 bc	0.326a	0.278abc	9.978abc
<i>S. sahendica</i> (1042)	1.150 bcde	0.689ab	0.460cd	1.525ab	0.393cde	3.943c	2.585d	6.563b	40.065bc	4.952c	0.329ab	0.186c	7.616c
<i>S. sahendica</i> (1043)	1.147 bcde	0.674ab	0.473c	1.447bcd	0.407bcde	3.834cd	2.655d	6.521b	40.972bc	5.400c	0.293ab	0.216bc	8.944bc
<i>S. sahendica</i> (1044)	1.069 cde	0.632b	0.437 cd	1.473abc	0.402cde	3.841cd	2.607d	6.481b	40.900bc	5.837c	0.307ab	0.261abc	9.523abc
<i>S. sahendica</i> (1045)	1.135 bcde	0.692ab	0.442cd	1.583ab	0.386cde	3.964c	2.517d	6.515b	40.010bc	5.450c	0.339ab	0.231abc	8.799bc
<i>S. sahendica</i> (7)	0.969 de	0.604bc	0.365d	1.692a	0.372e	4.015c	2.410d	6.464b	37.669c	6.284bc	0.378a	0.268abc	9.968abc
<i>S. sahendica</i> (8)	1.095 bcde	0.649ab	0.445cd	1.516ab	0.400cde	3.901c	2.618d	6.534b	40.630bc	5.420c	0.318ab	0.235abc	8.896 bc
<i>S. edmondi</i> (770)	0.949 e	0.518c	0.430cd	1.211e	0.450a	3.594e	2.940c	6.453b	45.399a	6.684bc	0.163d	0.273abc	11.306abc
<i>S. edmondi</i> (771)	1.396 a	0.764a	0.632a	1.218de	0.450a	3.562e	2.926c	6.499b	45.204a	5.583c	0.172d	0.232abc	10.038abc
<i>S. edmondi</i> (772)	1.278 ab	0.707ab	0.570ab	1.258cde	0.441ab	3.614ed	2.911c	6.540b	44.663a	5.732c	0.207cd	0.211bc	8.674bc
<i>S.kermanshahensis</i> (601)	1.064 cde	0.618bc	0.445cd	1.406bcde	0.413bcd	4.646a	3.305ab	7.994b	41.893ab	7.831abc	0.297ab	0.307abc	13.169ab
<i>S. khuzistanica</i> (705)	1.053 cde	0.636 b	0.416cd	1.579ab	0.385cde	3.904c	2.482d	6.429b	39.460bc	6.679bc	0.345ab	0.273abc	9.213bc

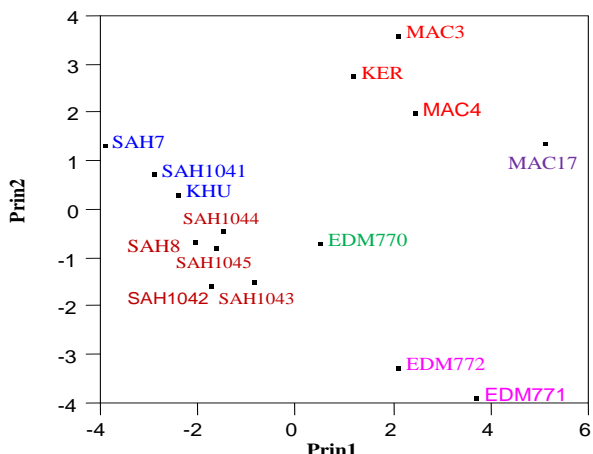


Fig. 2 Scatter plot of 15 populations for the first two principal components
 MAC: *S. macrantha*; EDM: *S. edmondi*; SAH: *S. sahendica*;
 KER: *S. kermanshahensis*;KHU: *S. Khuzistanica*

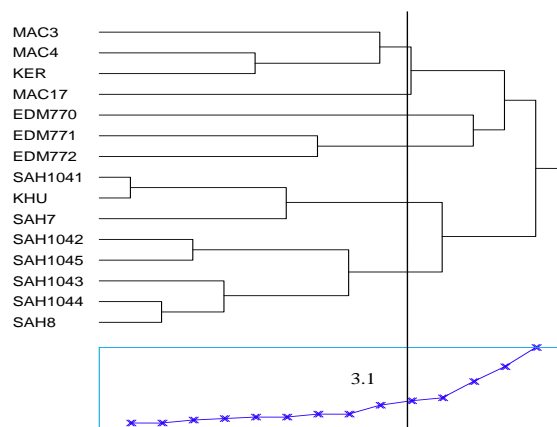


Fig. 3 Dendrogram of 15 populations of *Satureja* species by analyzing 11 karyotypic parameters using Ward's cluster analysis method. Cophenetic correlation $r=0.85$. MAC: *S. macrantha*; EDM: *S. edmondi*;
 SAH: *S.sahendica*; KER: *S. kermanshahensis*;KHU: *S. khuzistanica*

Table 5 Eigenvectors from the first two Principal components for 11 karyotype parameters to classify 15 populations of *Satureja* species.

Parameters	First component	Second component
TL	0.2798	-0.1831
LA	0.1842	-0.1287
SA	0.3257	-0.2063
AR	-0.3157	0.1713
CI	0.3103	-0.1831
LA%	0.1137	0.4215
SA%	0.3386	0.1706
RL%	0.2418	0.3393
TF%	0.3148	-0.1712
DRL	0.2134	0.3752
A ₁	-0.3072	0.1805
Eigen Value	7.0337	4.3456
Percentage of variance	50.2407	31.0403
Cum Percentage of variance	50.24	81.28

Table 6 Hierarchical Clustering, Method=Ward, Clustering History

Number of Clusters	Distance	Leader	Joiner
14	0.554562983	SAH1041	KHU
13	0.6121416236	SAH1044	SAH8
12	0.9576944508	SAH1042	SAH1045
11	1.0888036049	SAH1043	SAH1044
10	1.2424650466	MAC4	KER
9	1.2652000348	SAH1041	SAH7
8	1.5111192552	EDM771	EDM772
7	1.6540846005	SAH1042	SAH1043
6	2.5654195888	MAC3	MAC4
5	3.105471259	MAC3	MAC17
4	3.4513031617	SAH1041	SAH1042
3	5.2685470453	EDM770	EDM771
2	6.9404424054	MAC3	EDM770
1	8.9992772553	MAC3	SAH1041

DISCUSSION

In this study, chromosome numbers and detailed measurements of five native different species (15populations) of *Satureja* genus were examined in Iran. The concept of karyotype has been used to identify and differentiate chromosomes from different populations. Mitotic karyotype analysis is also useful in studying evolutionary problems.

Contrary to the report [20], *S. sahendica* species of the same origin that was declared to have the number of $2n=28$ chromosomes, this species has $2n=30$ chromosome along with its different populations. Also, in this report, chromosomal details such as karyotype formula and symmetry classes of Stebbins of two *S. khuzistanica* and *S. sahendica* species are different from similar collection areas reported by (Shariat *et al.* 2013) [22]. In order to refine the measure of karyotype asymmetry, we used the DI value, which can decode decipher even minor karyotype changes. The DI index plays an important role in sorting species within the same class of karyotype asymmetry in a specialty advancing order by allowing further grading, as illustrated by the arrangement of species within sections.

The results of variance analysis based on karyotypic features indicated the occurrence of quantitative variation in chromosome size of the studied populations. A significant effect of chromosomal traits proved karyotype changes between populations. This shows the importance of chromosome study to distinguish the state of evolution and affinity between different populations. This study showed that populations of a particular species also have diversity within themselves. The important factors in the separation of populations were LA%, SA%, RL% and DRL values. The present study shows the change in chromosomal traits as one of the mechanisms of inter and intra-species diversification in the *Satureja* genus, as well as the earlier cytological reports [20, 22, 30, 41-43].

In conclusion, from the examination of the chromosome number of the different species of *Satureja* it seems logical to conclude that the ploidy levels are different and the basic chromosome numbers should differ among species. Investigation of different populations of different *Satureja* species showed that because different populations grow in different places with varying climate, height, soil, slope etc, therefore, geographical effects can probably be effective on the classification of karyotypic traits on species and populations belonging to them. The results of the molecular markers used among the 15 populations also showed a high diversity as well as the amounts of components of essential oils (data not shown). These genomic differences could be used for breeding purposes. Thus, these studies could greatly help us in classification and taxonomic studies.

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REFERENCES

1. Nixon K. Genetic diversity in *Satureja* genus, diversity of life.org (DOL), Cornell University. 2006. from <http://www.Plantsystematics.org>.
2. Jamzad Z. *Thymus* and *Satureja* species of Iran, Research Institute of Forests and Rangelands, Tehran, Iran. 2009;1:171.
3. Sefidkon F., Jamzad Z., Mirza M. Chemical variation in the essential oil of *Satureja sahendica* from Iran. Food Chem. 2004;88:325-328
4. Sefidkon F, Jamzad Z. Essential oil analysis of Iranian *Satureja edmondi* and *S. isophylla*. Flavour Fragr. J 2006;21:230-233
5. Khadivi-Khub A., Salehi-Arjmand H., Hadian J. Morphological and phytochemical variation of *Satureja bachtiarica* populations from Iran. Industrial Crops and Products. 2014;54:257-265
6. Rechinger K. Flora Iranica, Labiatae, Akademische Druke- u.Verlagsanstalt. Graz Austria. 1982;150:479-480.
7. Farsam H., Amanlou M., Radpour M.R., Salehinia A.N., Shafiee A. Composition of the essential oils of wild and cultivated *Satureja khuzestanica* Jamzad from Iran. Flavour Frag J. 2004;19:308-310.
8. Hadian J., Mirjalili M.H., Kanani M.R., Salehnia A., Ganjipoor Chem P. Phytochemical and morphological characterization of *Satureja khuzistanica* Jamzad populations from Iran. Biodivers. 2011;8:902-915.
9. Kasyani Aval M., Tabaei Aghdaei S.R., Sefidkon F., Jafari A.A., Eftekhari S.A. Study the morphology and essential oil content in two *Satureja khuzistanica* populations under Tehran climatic condition. Annals of Biological Res. 2012;3:975-978.
10. Attar F., Einollahi N., Keyhani E., Keyhani J. Study on Superoxide Dismutase in *Satureja hortensis* L. Roots. Acta Hort. (ISHS). 2006;723:215-220.
11. Hadian J., Azizi A., Fakhr Tabatabaei M., Naghavi M.R., Jamzad Z., Friedt W. Analysis of the genetic diversity and affinities of different Iranian *Satureja* species based on SAMPL markers. Planta Med. 2010;76:1927-1933.
12. Bräuchler C., Meimberg H., Heubl G. New names in Old World *Clinopodium* L. - the transfer of the species of *Micromeria* sect. *Pseudomelissa* to *Clinopodium*. Taxon. 2006;55:977-981.
13. Bräuchler C., Meimberg H., Abele T., Heubl G. Polyphyly of the genus *Micromeria* (Lamiaceae) - evidence from cpDNA sequence data. Taxon. 2005;54:639-650.

14. Bräuchler C., Ryding O., Heubl G. The genus *Micromeria* (Lamiaceae), a synoptical update. *Willdenowia*. 2008;38:363-410.
15. Saidi M., Movahedi K., Mehrabi A.A., Kahrizi D. Molecular genetic diversity of *Satureja bachtiarica*. *Molecular Biology Reports*. 2013;40:6501-6508.
16. Zarei B., Kahrizi D., Sayfi T., Movahedi R., Ghaheiri M., Kazemi E. The Antibacterial Effect of Alcoholic Extraction of *Satureja bachtiarica* on Four Bacterial Human Pathogens. *Agric Biotechnology*. 2017;8:81-86.
17. Krogulevich R.E. Karyological analysis of the species of the flora of eastern Sayana. 19-48. In Malyshev L. I, Peschkova G. A, (eds.) *Fl. Prebaikalya*. Nauka, Novosibirsk. 1978.
18. Lövkvist B., Hultgård U.M. Chromosome numbers in south Swedish vascular plants. *Opera Bot*. 1999;137:1-42.
19. Markova M, Goranova V. Mediterranean chromosome number reports 5: 435-473. *Fl. Medit*. 1995;5:289-317.
20. Irani P., Hesamzadeh Hejazi S.M., Tabaei Aghdaei S.R. Karyological study on four species of *Satureja* (Lamiaceae) in Iran. *International J Biosciences*. 2014;4:229-240.
21. Markova M.L. Chromosome numbers of Bulgarian angiosperms. *Fitologija*. 1989;36:67-68.
22. Shariat A., Karimzadeh G., Assareh M.H. Karyology of Iranian Endemic *Satureja* (Lamiaceae) Species. *Cytologia*. 2013;78:305-312.
23. Boscaiu M., Riera J., Estrelles E., Güemes J. Números cromosómicos de plantas occidetales, 827-848. *Anales Jard. Bot. Madrid*. 2000;58:163-164.
24. Morton J.K. Chromosome numbers and polyploidy in the flora of Cameroon Mountain. *Opera Bot*. 1993;121:159-172.
25. Ferakova V., Murin A. In Index to chromosome numbers of Slovakian flora. Part 4. *Acta Fac. Rerum Nat. Univ. Comeniana, Bot*. 1974;23:1-23.
26. Reeves A., Tear J. *MicroMeasure* version 3.2. Colorado State University, USA. 2000. <http://www.colostate.edu/Depts/Biology/MicroMeasure/>
27. Hesamzadeh Hejazi S.M., Rasouli M. Cytogenetic study of some species of Vetch Genus (*Vicia* sp.) in Iran. *Iran. J Agric Sci*. 2006;37:213-225.
28. Javadi H., Hesamzadeh Hejazi S.M., Majnoon SH. B. Karyotypic studies of three *Thymus* (Lamiaceae) species and populations in Iran. *Caryologia*. 2009;62:316-325.
29. Hesamzadeh Hejazi S.M., Ziaei Nasab M. Cytogenetic study on several populations of diploid species of *Onobrychis* in natural gene bank of Iran. *Iran J. Rangelands for Plant Breed Genet Res*. 2009;16:158-171.
30. Hesamzadeh Hejazi S.M. Karyological study on three *Cicer* L. species (Fabaceae) in Iran. *Asian J Cell Biol*. 2011;6:97-104.
31. Ghasemi E., Hesamzadeh Hejazi S.M. Karyological studies in different populations of *Buxus hyrcana* (buxaceae) in Iran. *Iranian J botany*. 2018;24:156-162.
32. Soltanipoor M.A., Hesamzadeh Hejazi S.M., Jonoubi P. Karyotypic studies in eight populations of *Zhumeria majdae* Rech. f. & Wendelbo from Iran. *Caryologia*. 2017;70:222-228.
33. Huziwara Y. Karyotype analysis in some genera of compositae. V III, Further studies on the chromosome of aster. *Am. J. Bot*. 1962;49:116-119.
34. Romero Zarco C. A new method for estimating Karyotype asymmetry. *Taxon*. 1986;35:526-530.
35. Lavania U.C., Srivastava S. A simple parameter of dispersion index that serves as an adjunct to karyotype asymmetry. *J. Biosci*. 1992;17:179-182.
36. Stebbins G.L. *Chromosomal evolution in higher plants*. Edward Arnold publisher LTd, London. 1971;216pp.
37. Levan A.K., Sandberg A. Nomenclature for centrometric position on chromosomes. *Hereditas*. 1964; 52:201-220.
38. JMP. *JMP/STAT for windows*, version 3.1.2. SAS Institute Inc. 1995.
39. *STATISTI X.L*. *Statistical for windows*, version 1.8. University of Western Australia. 2007. from <http://www.statistixl.com>.
40. SAS. *SAS/STAT for Windows*. Version 6.12 SAS Institute Inc., Cary, NC. USA. 1996
41. Hesamzadeh Hejazi S.M., Ziaei Nasab M. Cytaxonomy of some *Onobrychis* (Fabaceae) species and populations in Iran. *Caryologia*. 2010;63:18-31.
42. Kalvandi R., Hesamzadeh Hejazi S.M., Atri M., Mirza M., Jamzad Z., Safikhani K. Karyotype analysis among 10 populations of *Thymus eriocalyx* species in Iran. *Ann. Biol Res*. 2012;3:3916-3925.
43. Salehi M., Hesamzadeh Hejazi S.M., Tabaei aghdaei S.R. Cytogenetic studies of two *Dracocephalum* (Lamiaceae) species and populations in Iran. *Int J Biosci*. 2014;4:100-108.