

Original Article

Sophora pachycarpa Root Extract has Ameliorative Effects on Testicular Injury but is Hepatotoxic in Carbon Tetrachloride Intoxicated Rats

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Article History

ABSTRACT

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Keywords

Carbon tetrachloride

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Background: Carbon tetrachloride (CCl₄) causes damages to the testicles, liver, kidneys, lungs and brain. In the present study, we investigated protective effects of *Sophora pachycarpa* Schrenk ex C.A.Mey. root extract on testicular damage, liver histopathology and sex hormone levels in CCl₄ intoxicated male rats.

Methods: In present study the rats were divided into 6 experimental groups (n=6): 3 groups as pre-treatment groups that received doses of 50, 100 and 250 mg/kg/days of *S. pachycarpa* extract respectively by gavage for 21 days before intraperitoneal injection of 500μl/kg CCl₄. CCl₄ group received 500μl/kg CCl₄ on the day of 21. Post-treatment group received 100 mg/kg/day *S. pachycarpa* for 10 days and 12h after injection of 250 μl/kg CCl₄. Control group received 1ml distilled water for 21 days. Liver and testis tissues were sampled for microscopic examination and sex hormone levels in the serum were evaluated.

Results: Treatment of *S. pachycarpa* root extract (250 mg/kg) lead to a considerable improvement in the histopathological changes caused by CCl₄ in the testis. However, the extract (250 mg/kg) caused inflammation and adverse tissue effects in liver. Serum levels of hormones also increased significantly in pre-treatment groups.

Conclusion: Based on the results, the *S. pachycarpa* extract can improve the changes caused by CCl₄ in the testis and sex hormones levels. But its higher doses can cause adverse effects in liver tissue.

INTRODUCTION

Carbon tetrachloride (CCl₄) is a clear, colorless, heavy, volatile and non-flammable liquid [1]. This chemical compound is an industrial solvent that is used for different purposes [1]. As an industrial material, CCl₄ is widely used in home cleaners, fire extinguishers, coolers, stimulants etc. Due to the harmful effects of carbon tetrachloride, it is important to control, prevent and eliminate its harmful effects [2,3]. Humans may be exposed to carbon tetrachloride through inhalation or oral [4].

CCl₄ causes damages to testes, kidneys, liver, brain and lungs through oxidative stress [5]. It induces oxidative stress by production of free radicals, trichloromethyl (CCl₃), trichloromethyldioxy (CCl₃ O₂) and other metabolites [6]. Oxidative stress means disturbance in the ratio of production of free radicals and oxidants of the body [7]. Free radicals are molecules, atoms or ions that contain one or

more unpaired electrons in the outer orbit of the molecules [8]. Free radicals are active chemicals that can rapidly react with cell components [9].

The liver is more vulnerable to oxidative stress than other organs [10]. CCl₄ is known as a liver toxin that can have effects such as fatty degeneration, fibrosis, hepatocellular death and carcinogenesis. In fact, CCl₄ is activated by cytochrome P450 to form the trichloromethyl radical (CCl₃). This radical can bind to cellular components (Nucleic acid, protein and lipid) [11].

Testicular toxicity had been known as the result of oxidative stress, caused by CCl₄ toxication. Testicular oxidative stress causes germ cells apoptosis, hormone regulation disorders and even male infertility [12]. Previous findings show that fruits and vegetables have a variety of nutrients and non-nutritive substances, which are called phytochemical compounds that can inhibit free

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radicals, interception the fatty oxidation and breaking of DNA, support the immune system by their antimicrobial, antioxidant and anticancer effects [13,14].

The *Sophora* plant belongs to Fabaceae family. It has 52 species reported worldwide, that three species are found in Iran, including *S. molis*, *Sophora alopecuroides* and *S. pachycarpa* (*S. pachycarpa*). The leaves of this plant are comprised, periodic and pinnate with a length of 10-18 cm [15]. The calyx has flowers similar to the flowers of the Pea family, white to yellow and fruits are clubshaped [15]. In Iran, *Sophora* can be found countrywide, especially in arid and semi-arid regions [15].

Study of chemical composition of *S. pachycarpa* has showed presence of alkaloids, flavonoids and steroid glucosides [16] which has anti-cancer, antibacterial, anti-inflammatory and antiviral activity [17]. In present study, the protective effects of *S. pachycarpa* root extract (SpRE) has been investigated in CCl₄ induced toxicity in male rats.

MATERIALS AND METHODS Extract Preparation

S. pachycarpa Schrenk ex C.A.Mey. plant was prepared from herbarium of the Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran. The roots of S. pachycarpa were placed in methanol at room temperature after drying. In order to prepare the crude methanolic extract, the extract filtered and solvent was evaporated at 40-45 °C. Then, for providing of required dosages, crude methanolic extract was dissolved in distilled water and KCl.

Animals

Male albino rats of Wistar strain (150-200g) were taken from Tabriz University of Medical Sciences. The rats were kept in polypropylene cages. The room temperature was kept at 20 ± 22 °C, with 40% - 60% relative humidity. Animals received standard food (pellets) and water. Rats were divided into 6 experimental groups, including 6 animals in each group.

Experimental Plan

Groups I, II and III as pre-treatment groups, received doses of 50, 100 and 250 mg/kg SpRE, respectively for 21 days by gavage. All three pre-treatment groups also received CCl₄ (500 μ l/kg)

intraperitoneally (ip) on the day of 21, 3-4 hours after the last dose administration of the extract [18]. Group IV was considered as control group that received 1 ml distilled water for 21 days.

Group V was considered as CCl₄ group that received CCl₄ (500 µl/kg) on the day of 21.

Group VI considered as post-treatment group that received 250 μ l/kg CCl₄ after 21 days and subsequently received 100 mg/kg SpRE for 10 days.

Histological Preparations

After the end of treatment period, animals were dissected. Testis and liver tissue samples were fixed in fixative (formalin 10%) for 48 h and then tissue slides were prepared by routine histotechnique method in order to light microscopic examination. Also, blood samples were taken for measuring serum levels of sex hormones by radio-immunoassay method.

Statistical Analysis

SPSS software (version 20) was used to analyze of the data. All results were analyzed using One-Way Variance test and Tukey's test (P< 0.05) and calculated as mean \pm standard error.

RESULTS

Hormonal Analysis

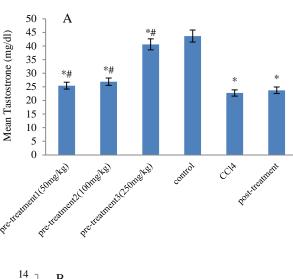
The data shown in table1 illustrate that follicle stimulating hormone (FSH) levels were markedly (P< 0.05) higher in pre-treatment III (8.86 ± 0.06) and post-treatment groups (6.64 ± 0.41) in comparison to CCl₄- group (5.54 ± 0.5) (Fig. 1).

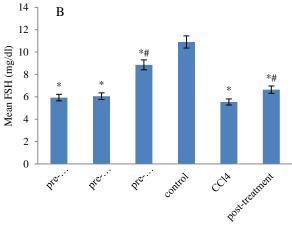
Also, the mean concentration of luteinizing hormone (LH) was significantly increased (P< 0.05) in pre-treatment I (28.38 \pm 0.57), II (29.15 \pm 0.45), III (31.76 \pm 0.44) groups and post- treatment group (24.32 \pm 0.33) in comparison with CCl₄- group (16.98 \pm 0.22) (Fig.1).

Testosterone levels in pre-treatment I (25.44 \pm 0.14), pre-treatment II (26.88 \pm 0.28) and pre-treatment III (40.28 \pm 0.23) groups has increased significantly (P< 0.05) compared to CCl₄- group (22.74 \pm 0.4) (Fig. 1).

Histopathological Studies

According to histologic examination, spermatogenic cells are limited to the lumen of the seminiferous tubular basement membrane in CCl₄- group (Fig. 2.A).





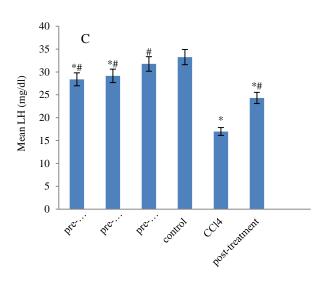
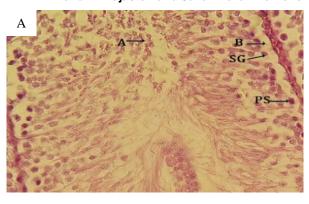
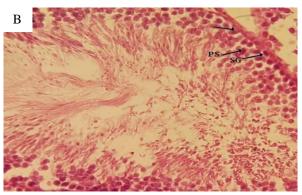


Fig.1 Comparison of the mean serum concentrations of LH, FSH and testosterone among the studied groups. The significant difference (P< 0.05) of the experimental groups compared to the control group is shown with * and the significant differences (P<0.05) of the treatment groups compared to CCl₄ group is shown with #.





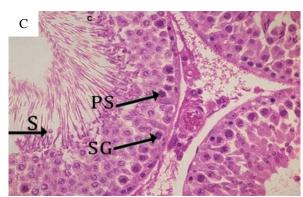
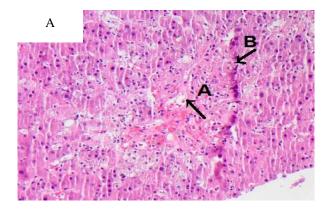
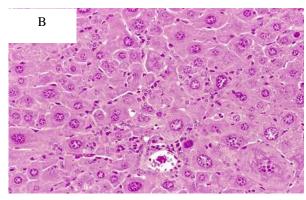


Fig. 2 A. Section from seminiferous tubules of CCl4 group. A: loss of spermatogenic cells inside the lumen; B: development of space between the basement membrane and spermatogenic cells (H&E stain, ×400). B. Seminiferous tubular section of SpRE group 250 mg/kg + CCl4 (H&E stain, ×400). C. Section from seminiferous tubule of control group. S, Spermatid; PS: primary spermatocyte; SG: spermatogonia (H&E stain, ×400).

Administration of CCl₄ caused destruction in germinative epithelium of seminiferous tubules, loss of germ cells and falling of cells into the lumen. Also, nuclei of spermatogenic cells were crumpled and the cells were out of order. There was loss of intercellular connections. Also results showed that at pre-treatment groups SpRE moderated the effects of CCl₄, especially at higher doses (250 mg/kg). Liver histopathological examination of normal group revealed normal distinct cells, portal spaces and hepatocytes. In comparison, pre- treatment group I (50 mg/kg) showed tissue damage. There

was cell swelling and karyomegaly in pre-treatment group II (100mg/kg). As well as inflammation, cell swelling, and cell death were illustrated in pre-treatment group III (Fig. 3).





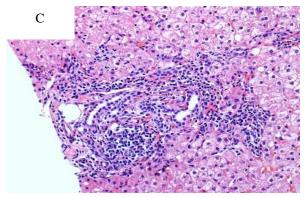


Fig. 3 Liver Section. A. Necrosis in pre-treatment group I. A: expansion of sinusoids, B: Apoptotic bodies. B. Karyomegaly of hepatocytes in pre-treatment II. C. Hepatitis in pre-treatment group III. (H&E stain, × 400).

DISCUSSION

Chloride hydrocarbons are industrial or organic solvents. These hydrocarbons are used in chemical industry, medical and other fields [19]. Carbon tetrachloride that penetrates the body through the skin and inhalation cause damage to organs including testis and liver [2,3].

The testis is one of the organs that carbon tetrachloride causes damage through oxidative stress [12]. Oxidative stress leads to germ cells apoptosis;

subsequently causes hypo-spermatogenesis and endocrine signaling. Therefore, oxidative stress has been reported as one of the factors of male infertility [20].

The regulation of hepatocytes by thyroid hormones has indicated that there is a clear correlation between the liver and thyroid hormones [21]. Hyperthyroidism significantly alters the gonadotropin and prolactin axis and severely affecting sperm function [22].

The liver is known as one of the organs affected by carbon tetrachloride. In fact, carbon tetrachloride is known as one of the reasons of hepatic-cirrhosis as a result of reducing the levels of antioxidant enzyme activities such as catalase, peroxidase, superoxide dismutase and etc. [23].

Medicinal plants have been used for treatment of many diseases in the recent decades, due to cheapness, availability and to be effective [24]. Plants are known as a rich source of antioxidant compounds such as flavonoids. Studies have shown that the use of plants containing antioxidants has a significant relationship with the reduction of human diseases related to oxidative stress [25].

In this study, CCl₄ reduced levels of the sex hormones in comparison with control group. Therefore, it can be said that CCl₄ can cause infertility in men. However, *S. pachycarpa* root extract as a medicinal plant led to increase the sex hormones levels contrasted to CCl₄ group, especially in pre-treatment group III. So this medicinal plant can be effective in preventing infertility caused by CCl₄[26].

In a similar study, moderating effects of various fractions of *Rumex hastatus* roots in CCl₄- induced testicular and hepatic damage has been shown [26].

Histopathological examination in present study was indicated that administration of SpRE before exposure to CCl₄ (especially in 250 mg/kg) improves changes caused by CCl₄ in testis. However, SpRE aggravated the toxic effects of CCl₄ in the liver of male rats. Therefore, SpRE may has anti-cancer properties so that lead to hepatotoxicity. It seems that apoptosis caused by dichloromethane and *Sophora* flavonoids G are involved in this toxicity; therefore, caution is required in its use [27]. It was demonstrated that *Rumex hastatus* root extract can improve liver disorders induced by CCl₄ such as necrosis, inflammatory cells and ballooning

of cells. This extract improved the disorders induced by CCl₄ in the testis and liver [26].

The study of Xiaohe and *et al.* showed that administration of Byakangelicine (100 mg/kg) after four weeks through oral gavage until the end of eight weeks modified disorders such as fibrosis, hepatocyte apoptosis and stellate cell inhibition caused by carbon tetrachloride in liver [28].

In other study, it has showed that after treatment with Propolis (200 mg/kg), there were improvement in hormone levels and spermatogenesis. This may be due to the antioxidant properties of the extract [5].

Aramjoo and *et al*, stated that *S. pachycarpa* seed (300mg/kg) has flavonoids such as polyohenols, that lead to improvements of renal toxicity and reproductive disorders [29]. They showed that *S. pachycarpa* seed extract at dose of 300 mg/kg can improve centrilobular necrosis, fatty change and infiltration of inflammatory cells in CCl₄ intoxicated rats that is contrary to the results of the present study [29].

A similar study on carbon tetrachloride-induced hepatotoxicity and hypogonadism in rats indicated moderating effects of Rutin (vitamin P) on toxicity. Rutin contains quercein flavonoids and disaccharide rutinose [30]. It has been found that Rutin prevents liver damage caused by methotrexate in rats [31]. Also, Rutin has a promising efficacy on infertility induced by oxidative stress in males [32]. In study by Elsawy H and et al, in CCl4 group showed centrilobular necrosis, hepatocyte degeneration, vacuolar fatty alteration and slight mononuclear cell infiltration [33]. tetrachloride affected spermatogenesis. In this way, the serum levels of testosterone, LH and FSH were significantly reduced compared to the control group. Carbon tetrachloride decreased the secretion of hormones in rats directly through the central nervous system and Leydig cells or indirectly hypothalamic-pituitary the through receptor axis [34].

Elsawy and *et al.*'s research showed that high levels of serum cholesterol are related to testicular disorders and decreased sperm quality. Therefore A diet high in antioxidants and low in cholesterol improves spermatogenesis [35].

In another study by Badr and Colleagues was indicated that carbon tetrachloride (3ml/kg) poisoning caused hypospermatogenesis in irregular

seminiferous tubules with primary edema in the intestitial spaces in male rats. But the administration of Parsley Leaf (*Petroselinum crispum*) in a dose of 0.5 ml/kg moderated these changes. Also, Parsley Leaf treatment induced Significant increase in serum levels of Testosterone, LH and FSH. This improvement was due to the presence of natural antioxidants in the extract [12].

CONCLUSION

This study determined that exposure to CCl₄ in male rats damages the liver and testis. Due to presence of various flavonoids, the root extract of *S. pachycarpa* can improve the harmful effects of the carbon tetrachloride in testicular tissue and the levels of sex hormones. Therefore, it can be said that SpRE is useful in preventing infertility caused by CCl₄. However, SpRE led to cytotoxicity in intact hepatic cells.

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Conflict of Interest

The authors declare no conflict of interest.

REFERENCES

- 1. Bahashwan S., *et al.* cronic mitigates carbon tetrachloride-induced liver toxicity in rats. J. Taibah University Medical Sci. 2015; 140-149.
- 2. Al Amin A., Menezes R.G. Carbon tetrachloride toxicity. StatPearls. 2020.
- 3. Torres-González L., Muñoz-Espinosa L.E., Rivas-Estilla A.M., Trujillo-Murillo K., Salazar-Aranda R., De Torres N.W., *et al.* Protective efect of four Mexican plants against CCl4-induced damage on the Huh7 human hepatoma cell line. Ann Hepatol. 2016;10(1):73–9.
- 4. Manibusan M.K., Odin M., Eastmond DA. Postulated carbon tetrachloride mode of action: a review. J Environ Sci Health C. 2007;25(3):185–209
- Hashem A.S. Defensive impact of propolis against CCl4 actuated rats testicular damage. J Adv Vet Anim Res. 2021; 8:70-77.
- Unsal V., Cicek M., Sabancilar I. Toxicity of carbon tetrachloride, free radicasl and role of antioxidants. Rev Environ Health. 2020; 36: 279-295.
- 7. Yang al *et al.* The Role of Oxidative Stress and Natural Antioxidants in Ovarian Aging: A Review. Front Pharmacol. 2021; 11: 1-18.

- 8. Di Meo S., Venditti P. Evolution of the Knowledege of Free Radicals and Other Oxidants. Oxid Med Cell longev. 2020; 1-32.
- 9. Sydney J. Aprioku. Pharmacology of free radicals and the impact of Reactive oxygen species on the testis. J. Reprod infertil. 2013; 14: 158-172.
- 10. Sadasivam N., *et al.* Oxidative stress, Genomic integrity and Liver Diseases. Molecules., 2022, 27.
- 11. Lutz W D Weber et al. hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicologinal model: A Review. Crit Rev Toxicol. 2008; 33: 105-136.
- 12. Gehan M., et al. Antioxidant Potential of Parsley Leaf (Petroselinum crispum) essential oil on hypothyroidism and testicular injury in mice intoxicated by carbon tetrachloride. Biomed Res Int. 2021.
- 13. Harish Nayaka M.A., Sathisha U.V., Dharmesh S.M. Cytoprotective and antioxidant activity of free, conjugated and insoluble-bound phenolic acids fromswallow root (Decalepis hamiltonii). Food Chem. 2010; 119: 1307-12.
- 14. Shukla S., Mehta A., Bajpai V.K., Shukla S. In vitro antioxidant activity and total phenolic content of ethanolic leaf extract of Stevia rebaudiana Bert. Food and Chemical Toxicology. 2009; 47: 2338-43.
- 15. Eisenman S.W., Zaurov D.E., Struwe L. Medicinal plants of Central Asia: Uzbekistan and Kyrgyzstan. New York: Springer. 2013; pp.203-60.
- Muosavi SH., et al. In vitro Evaluation of Cytotoxic and Apoptogenic Properties of Sophora pachycarpa: Iranian J. Pharmaceutical Res. 2014; 2: 73-665.
- 17. Es.Haghi M etal. Protective effects of Cornus mas fruit extract on carbon tetrachloride induced neohrotoxicity in rats. 2014; 24(5):291-296.
- 18. Kumar Sh. Chemistry and Biological Activities of Flavonoids: An Overview. The Sci World J. 2016; 16: 1-16.
- 19. Meng Y., *et al.* Enhanneed degradation of carbon tetrachloride by surfactant-modified zero-valant iron. J Zhejiang Univ Sci B, 2006; 7: 702-707.
- 20. Turner T., lysiak J. oxidative stress: a common factor in testicular dysfunction. J. Andrology. 2008; 29: 488-489.
- 21. Malik R., Hodgson H. The relationship between thyroid gland and liver. QJM. 2002; 95: 559-569.
- 22. Abalovich M., *et al.* Hypothalamic-Pituitary-Testicular axis and seminalo parameters in hyperthyroidism males. Thyroid. 1999; 9: 857-63.
- 23. Sajjad M., *et al.* Phytochemical, antioxidant and hepatoprotective effects of Alnus nitida bark in carbontetrachloride challenged Sprague Dawley rats.

- BMC Complementary and Alternative Medicine. 2016; 16, 268
- 24. Roy A., et al. in vitro antioxidant and free radical scavenging activity of the ethanolicextract of dioscoreavillosa (wild yam) tubers. Drug Invent. 2011; 3: 214-5.
- 25. Halliwell B., Gutteridge J. Cellular responses to Oxidative stress adaptation damage, repair, senescence and death. Free radicals in Biology and Medicine. NewYork: Owford University Press. 2015.
- 26. Sahreen S., Rashid Khan M., Ali Khan R. Ameliorating effect of various fractions of Rumex hastatus roots against hepato- and Testicular toxicity caused by CCl4. 2013.
- Banan Khojasteh S.M., Javanmard Khameneh R. Sophora pachycarpa root extract improves testicular damage in carbon-tetrachloride intoxicated rats. 2018; 20(8):e 64950.
- 28.Xiaohe Li., *et al.* Byakangelingelicine protects against carbon tetrachloride-induced liver injury and fibrosis in mice. J Cell Mol Med. 2020; 24: 8623-8635.
- Aramjoo H., et al. Protective effect of Sophora pachycarpa seed extract on carbon tetrachloride-induced toxicity in rats. BMC Complementary Medicine and Therapies. 2022; 22: 76-88.
- Ganeshpurkar A., Saluja A.K. The pharmacological potential of rutin. Saudi Pharmaceutical J. 2017; 25(2):149_164.
- 31. Erdogan E., Ilgaz Y., Gurgor P.N., Oztas Y., Topal T., Oztas E. Rutin ameliorates methotrexate induced hepatic injury in rats. Acta Cirurgica Brasileira. 2015; 30(11):778-784.
- 32. Mehfooz A., Wei Q., Zheng K., Fadlalla M.B., Maltasic G., Shi F. Protective roles of Rutin against restraint stress on spermatogenesis in testes of adult mice. Tissue and Cell. 2018; 50:133-143.
- Elsawy H., Badr G.M., Sedky A., Abdallah B.M., Alzahrani A.M., Abdel-Moneim AM. Rutin ameliorates carbon tetrachloride (CCl4)-induced hepatorenal toxicity and hypogonadism in male rats. 2019; PeerJ 7:e7011.
- 34. Okolo K.O., Siminialayi I.M., Orisakwe O.E. Protective effects of Pleurotus tuberregium on carbon-tetrachloride induced testicular injury in Sprague Dawley rats. Frontiers in Pharmacology. 2016; 7:Article 480.
- 35. Khorrami A., Ghanbarzadeh S., Ziaee M., Arami S., Vajdi R., Garjani A. Dietary cholesterol and oxidised cholesterol: effects on sperm characteristics, antioxidant status and hormonal profile in rats. Andrologia. 2015; 47(3):310-317.